Association Between moleculars and Osteoporotic Fracture Risk : A systematical review

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Abstract

Osteoporosis is a systemic chronic skeletal disease, which is characterized by low bone mineral density (BMD) and increased risk to osteoporotic fractures (OFs). OFs are associated with high mortality and morbidity, and seriously affect the life quality of patients. Osteoporosis is prevalent in the middle-aged and elderly population, especially the postmenopausal women. With population aging, osteoporosis becomes a world-wide serious public health problem. Early recognition of the high-risk population followed by timely and efficient intervention and/or treatment is important for preventing OFs. In light of the high heritability and complex pathogenesis of OP, comprehensive consideration of significant biological/biochemical factors is necessary for accurate risk evaluation. For this purpose, we reviewed recent research progress on moleculars which are diagnostic and/or predictive of OFs risk. Future integrative analyses and systematic evaluation of these moleculars may facilitate developing novel methodologies and/or test strategies, i.e., biochips, for early recognition of osteoporosis, hence to contribute to preventing OFs in the world.

Graphical Abstract

Osteoporosis, which is characterized by low bone mineral density (BMD) and increased risk to osteoporotic fractures (OFs), is prevalent in the middle-aged and elderly population, especially in the postmenopausal women. We focused on several types of important molecules, including proteins/peptides, RNAs, lipids, to gain comprehensive understanding and to generate novel perspectives in predicting and diagnosing OFs.

Keyword: Osteoporotic fracture, molecular, Predictive, Diagnostic, Risk factor

Introduction

Osteoporosis is a chronic skeletal disease, which is characterized by low bone mineral density (BMD) and high risk to osteoporotic fracture (OF). Osteoporosis is prevalent in the middle-aged and elderly population, especially in the postmenopausal women. With population aging, osteoporosis and OFs have become an increasingly serious public health problem. A report from World Health Organization (WHO) in 2015 noted that OFs place a heavy burden on individuals and have important health consequences in both developed and developing countries [1].

A commonly used method to predict fracture risk is the FRAX tool [2], which has been modified or simplified in various countries and regions. Since the method was released, no molecular information has been incorporated into the prediction model thus far. A molecular, which was discovered by clinical test, is mainly composed of proteins, followed by small molecules and cells, and is measured as an indicator of normal biological or pathogenic processes, or a response to an exposure or intervention(FDA-NIH: molecular-Working-Group,2016).

The 10-year risks to hip fracture and major OF, which were calculated using the Chinese FRAX® model by the National Osteoporosis Foundation (NOF), suggested that the risk to OF increases with age in Chinese[3]. A recent report showed that the FRAX method dramatically underestimate the fracture risk of Chinese population [4], which calls for more significant factors to be incorporated and more fine-tuned method to be applied for Chinese population for the purpose of precision prediction/medicine.

Osteoporosis results from imbalanced bone remodeling favoring bone loss due to relatively increased bone resorption by osteoclasts and/or decreased bone formation by osteoblasts. OFs are determined by both genetic and environmental factors, as well as their interactions. As reviewed previously, a large number of genes have been identified to be associated with BMD variation and OFs in humans [5, 6]. Meanwhile, rapid progress has been made in identification of novel biomolecules associated with OFs risks [7]. Therefore, it is reasonable for us to believe that targeted evaluation of a panel of significant biomolecules would become feasible in the near future, and facilitate developing effective moleculars to be applied for OF risk prediction.

For the above purpose and perspective, here we reviewed recent research progress on moleculars diagnostic and/or predictive of OFs, as follows.

Methods

Since 1979, the number of publications on OF moleculars increases dramatically year by year, showing that continuous attention has been cast from investigators to the research field. To obtain a comprehensive collection of publications on moleculars of OFs, we performed an unrestricted PubMed search specifying co-occurrence of the terms "osteoporotic fractures and molecular", "osteoporotic fractures and protein", "osteoporotic fractures and RNA", "osteoporotic fractures and metabolite", "osteoporotic fractures and lipid", then collected 15,781 publications.

To gain a comprehensive understanding and to generate novel perspectives, we reviewed the research progress in moleculars diagnostic and/or predictive of OFs. Considering that quite many reviews have already summarized the research progress at gene/SNP level, e.g., from genome-wide association and meta-analyses [5, 6], in this review, we focused on summarizing the progress on other types of molecules, such as proteins/peptides, RNAs, lipids, etc. As follows, the predictive and diagnostic moleculars were categorized and presented, respectively. However, this review based on the population of epidemical research, further studies and trials are needed to demonstrate the clinical values and the possible to be target of these novel moleculars.

Potential predictive moleculars of OFs

Generally speaking, recent studies identified many circulating moleculars for OFs in the elderly. A majority are proteins which are predictive of OFs[8], such as insulin-like growth factor (IGF), periostin (POSTN), etc. In addition, more and more evidences suggest that metabolites, including nucleic acid, amino acid and lipid mediators, could predict OFs risk, as well. To date, further clinical application for these novel moleculars should be explained in clinical trials.

1. Protein and peptide moleculars

A dozen of predictive protein moleculars associated with incident OF have been identified from prospective studies listed in **Table 1**. Risk predictive protein moleculars, including IGF binding protein-1(IGFBP-1)[9], periostin [10, 11], adiponectin[12, 13], C-reactive protein (CRP) [14], bone-specific alkaline phosphatase (BAP) [15] and fibroblast growth factor 23 (FGF23) [16, 17], were all positively associated with incident OFs.

Lundin H *et al.* discovered that IGF binding protein-1 (IGFBP-1) protein level in serum was positively correlated with the risk of hip fractures [Adj. hazard ratio(HR)=1.46 (1.08-1.99) per SD increase] and major OFs [HR=1.33 (1.05–1.69), P=0.017] in a population of 351 Sweden

women [9]. Periostin is a secreted γ -carboxyglutamic acid-containing protein, which is expressed mainly in the periosteum of adult individuals[18]. Higher level of periostin in serum increases the fracture risk [10, 11]. For instance, Rousseau JC et al. discovered that the highest quartile of serum periostin was associated with an increased risk of incident fracture in a prospective study of 607 postmenopausal women [relative ratio (RR)=1.88 (1.1-3.2)] after adjustment for age, fragility fracture, and hip BMD T-score[10]. Periostin fragment resulting from cathepsin K digestion predicted OF risk in females as well [HR=2.14(1.54-2.97)] [11]. In 2012, Johansson H et al. found that the OF risk increased in parallel with increasing serum adiponectin [HR/SD=1.46(1.23-1.72)] and persisted after multivariate-adjusted analysis [HR/SD=1.30(1.09-1.55)] in 999 elderly Swedish males[12]. In 2014, Johansson H et al. highlighted the predictive value of serum adiponectin for OF in males [HR=1.47(1.23-1.77) per SD after adjusted for age and time since baseline, and HR=1.40(1.16-1.68) per SD after adjusted for femoral neck BMD and age][13].

Besides, the serum CRP level was associated with a risk to any type of OFs [HR=1.06(1.02-1.11)], hip fractures (HF) [HR=1.09(1.02-1.17)] and vertebral fractures (VF)[OR=1.34(1.14-1.58)] in a prospective study of 6,386 participants (59% female) [14]. However, the clinical value of CRP and its potential diagnostic ability for OF need to be further validated, especially in single patient, since we only explained its associations based on epidemical research. Serum BAP was also reported to predict an increased risk of osteoporotic VF [RR per SD=4.38(1.45-13.21)] in 65 postmenopausal women with years of menopause (YSM) <5 years. In 457 postmenopausal women with $YSM \ge 5$ years, the upper tertile for BAP had a greater risk of vertebral fractures[RR=1.39(1.12-1.74)][15]. Furthermore, Mirza MA et al. pointed that FGF23 protein level in serum above 55.7 pg/mL was associated with an increased risk to HF [HR=2.30(1.16-4.58)] and non-VF [HR=2.163(1.01-2.63)], respectively in a population-based OF study in men (MrOS; N=2868), and subjects in the highest quartile of serum FGF23 protein level (>57.4 pg/ml) were at a 63% increased risk to non-vertebral OFs, compared to those in the lowest three quartiles [16]. Lane NE et al. discovered that in men with poor renal function only (eGFR_{crCvs}< 60ml/min/1.73m²), the relative hazards (RH) of hip and non-spine OFs was 2.02 (95% CI=1.07-3.79) in the highest quartile of serum FGF23 protein level, as compared to the rest [17].

Besides, a couple of protective protein moleculars for OFs, including sclerostin [19] and the endogenous secretory receptor for Advanced Glycation End Products (esRAGE)-to- Pentosidine (PEN) ratio [20], have been identified thus far. For example, Szulc Pet al. reported that French men with higher concentration of serum sclerostin had lower OFs risk [HR \square = \square 0.55(0.31-0.96),

 $P \square < \square 0.05$ adjusted for hip BMD][19]. In addition, theesRAGE was associated with reduced activity of PEN [20]. Tamaki J *et al.* found that the esRAGE-to-PEN ratio was negatively correlated with incident OFs in 1,285 elderly male Japanese [(HR (95%Cl)= 0.67 (0.45-0.98) and 0.64 (0.43-0.95), adjusted for age, glycated hemoglobin A1c, and T-score of BMD at FN or LS] [20].

As for the peptide moleculars, such as bone resorption markers CTX [21-23] and bone formation marker PINP [23, 24], showed great predictive values for OFs. For example, Early in 1996, Garnero P *et al.* found that CTX level above the upper limit of the premenopausal range was associated with an increased risk of hip OF in a population of 7,598 healthy French [OR=2.2(1.3-3.6)][21]. In 2002, Garnero P *et al.* found that women with both increased urinaryalpha-L/beta-L-CTX ratio and low femoral neck BMD (T score<-2.5) had a higher risk of OFs[RR=4.5(2.0–10.1)] in a prospective study [22]. Yoshimura N *et al.* reported that serum beta-CTX and NTX was significantly related to the occurrence of spinal OFs in Japanese women (beta-CTX: HR=1.80, *P*<0.001; NTX: HR=1.96, *P*<0.01) after adjusting for confounders in multivariate analysis [23], and serum PINP level was significantly higher in OFs than controls in both men (HR=2.80, *P*<0.05) and women (HR=1.65, *P*<0.05) [23].

2. Potential Metabolite moleculars

Metabolites play an essential role in predicting OFs. Some polar metabolites are significantly involved in nucleic acid metabolism, amino acid metabolism and lipid metabolism, which are closely related to the imbalance between bone resorption and formation and may underlie the development of OFs [25]. Predictive metabolite moleculars associated with OFs risk were summarized in **Table 2** and presented in the following three categories.

2.1 Nucleic acid metabolite

Some nucleic acid metabolite, play a key role in the bone metabolism. According to the EPIDOS cohort study including 7,598 female French, high level of urine free deoxypyridinoline was associated with increased risk of HFs, and free deoxypyridinoline secretion above the upper limit of the premenopausal range was also associated with an increased HFs risk [OR=1.9(1.1-3.2)][21]. Yoshimura N *et al.* found that high level of urinary deoxypyridinoline

> significantly predicted 10-year OFs risk at the lumbar spine in women (HR=1.40, P<0.05) but not in men [HR=1.53(0.63-3.73)][23]. To be honest, the significant associations were only validated in epidemical research, rather than in specific clinical treatment, especially for single patient, so the potiential predictive value of them needs to be addressed.

2.2 Amino acid moleculars

Plasma total homocysteine(tHcy) was frequently reported to be associated with OFs risk [26, 27]. The adjusted HR (95% CI) for OFs in subjects with high (>=15 microM) vs. low levels (<9.0 microM) of tHcy was significant in women [HR=2.42 (1.43-4.09)] but not in men [HR=1.37 (0.63-2.98)] ina prospective study of 4,776 elderly [26]. Whereas incidence of OFs was associated with quartiles of homocysteine (Hcy) (P=0.047), higher risks of OFs was observed in females aged 55 years and over [adjusted for age, BMI, FN-BMD, HR=1.82(1.02-3.22)] from the Rotterdam Study (RS) in a prospective study of 7 following-up years[27].

Dimethylglycine (DMG) is a product of the choline oxidation pathway and generated from betaine during the folate-independent remethylation of Hcy to methionine. Lower level of plasma DMG was associated with an increased risk of incident HF [HR=1.70(1.28-2.26)], as compared to the highest tertile[28].

2.3 Lipid mediators

Osteoporosis or osteopenia often co-exist with lipid metabolism disorders [29]. Studies suggested that peroxisome proliferator activated receptor gamma (PPAR γ) may play a key role in the pathogenesis of both osteoporosis and lipid metabolism disorder, and lipid metabolism disorders caused by bone marrow fat may be the main cause of bone marrow microcirculation disorders and osteopenia [30].

Sphingosine-1-phosphate (S1P) is a lipid mediator which acts through S1P receptors.S1Pplays diverse roles in cell differentiation, apoptosis, proliferation, motility, bone metabolism, and the most prominent role seems to be the augmentation of bone resorption [31]. In another prospective longitudinal study of 707 postmenopausal women, the plasma S1P levels were significantly higher in the women with incident OFs than in those without osteoporosis related fractures (ORFs) (7.23±0.79 μ mol/L vs. 5.02±0.51 μ mol/L; *P*<0.001) [32].After adjustment for age and other confounders, the HR of OFs was 6.12 (95% CI: 4.92-7.66) per SD increase in plasma S1P level

and the women in the highest quartile of S1P levels had a significantly increased fracture risk [HR=9.89(2.83-34.44)][32].

Ginsberg C et al. revealed the association of vitamin D with hip BMD and the risk of HF among 890 participants followed-up for ~8.4-years, both higher 24,25(OH)₂D and vitamin D metabolite ratio (VMR, i.e., the ratio of 24,25(OH)₂D to 25-hydroxyvitaminD (25OHD)) were associated with lower risk of HF [HR=0.73(0.61-0.87)] and 0.74 (0.61-0.88) per SD increase, respectively] [33]. Swanson CM et al. also reported that 25OHD and 1,25(OH)₂D were protective against HFs [34]. However, previous studies suggested that the hip fracture protection found with higher 250HD levels is largely mediated by BMD[35]. Specifically, the risk of HFs was higher in men with the lowest 1,25(OH)₂D levels (8.70 to 51.60 pg/mL) after adjustment for baseline hip BMD [HR (95% Cl)=1.99(1.19–3.33)]; and significantly increased risk of HFs was also observed for the lowest quartile of 25OHD (3.13 to 20.90 ng/mL) in American man [HR=2.01(1.24-3.26)][34]. However, there existed some inconsistent results regarding to the associations between 25(OH)D and high risk of HF[36]. To our knowledge, no prior study has evaluated whether 24,25(OH)₂D concentrations or the VMR are more strongly associated with bone health than 25(OH)D alone. Additionally, the associations of 25OHD and 1,25 (OH)2D with baseline BMD and BMD change were independent of each other. These results do not support the hypothesis that measures of 1,25(OH)2D improve the ability to predict adverse skeletal outcomes when 25OHD measures are available. Still, there is long way before reaching the potential diagnostic role of 25OHD, 1,25(OH)₂D and risks to OFs. Epidemical outcomes need to be further elaborated when apply to clinical trials.

Besides, Harris TB *et al.* reported that higher concentration of plasma polyunsaturated fatty acids (PUFAs) was associated with lower OFs risk in older adults [37]. Among 1,438 participants, 540 participants with incident OFs, the highest tertile of plasma PUFAs was associated with decreased OFs risks [HR=0.60(0.41-0.89)] in men; PUFAs tended to be inversely associated with OFs risks in women ($P_{trend} = 0.06$)[37]. However, PUFAs are novel moleculars related to OFs and only conducted in epidemcal research among population, the further significant value in clinical usage to treat OFs need to be validated in single patient.

Potential Diagnostic moleculars of OFs

Some moleculars, such as insulin-like growth factor-1 (IGF-1), and IGF-binding protein-3 (IGFBP-3), etc. demonstrated great diagnostic values for OF. Recently, more scientific research suggested that microRNAs (miRNAs) and metabolites are diagnostic of OFs risk, as well.

1. Potential protein moleculars

Potential diagnostic protein moleculars associated with OF risks were identified in cross-sectional studies, which were summarized and listed in **Table 3**. In 1997, Sugimoto T *et al.* reported that serum IGF-I protein level was positively correlated with BMD and negatively associated with spinal OF in Japanese women [38]. Nakaoka D *et al.* also discovered that IGF-I protein was negatively correlated with the risk of spinal OFs in Japanese postmenopausal women [HR=0.19(0.07-0.59)][39]. Similar results were reported by Yamaguchi T *et al.* for VF [OR=0.29 (0.15-0.57) per SD increase, P=0.0003] in Japanese postmenopausal women [40]. Besides, IGF-binding protein-3 (IGFBP-3) was also negatively associated with both spinal OFs (r=-0.409, P<0.0001) [38] and VF [OR=0.31(0.16-0.61), P=0.0007][40] in Japanese women. As reported by Kim BJ *et al.*, plasma periostin was a risk factor of any fracture [OR=1.50(1.14-1.97), P=0.003], non-VF fracture [OR=1.59(1.12-2.24), P=0.009]in Koreans [41]. The odds for non-VF was 2.48-fold higher in the highest vs. lowest tertiles of plasma periostin level (95% Cl=1.10-5.61)[41]. Consistently, Yan *et al.* reported that serum periostin level was positively correlated with femoral neck BMD (r=-0.529, P<0.001) in Chinese elderly females[42], supporting that it is a risk marker of OFs.

2. Potential MicroRNA moleculars

MicroRNA (miRNAs) is a class of small (~22 nucleotides), single-stranded noncoding RNAs. They are considered "micro-managers" of gene expression, and their role in regulating biological functions has been increasingly recognized. The diagnostic miRNAs moleculars associated with OFs risk were summarized in **Table 4**.

Early in 2014, Seeliger C *et al.* identified 9 serum miRNAs (miR-21, miR-23a, miR-24, miR-93, miR-100, miR-122a, miR-124a, miR-125b, and miR-148a) significantly up-regulated in OFs in a case-control study, which presented modest diagnostic performance individually (AUC=0.61-0.77, P<0.05). Among the above nine miRNAs, miR-21, miR-23a, miR-24, miR-100 displayed a significantly higher expression level in bone tissues in OFs compared with controls [43]. In 2015, Panach L*et al.* showed that 3 miRNAs in serum (miR-122-5p, miR-125b-5p and miR-21-5p) were up-regulated in fracture cases vs. controls [44]. Indeed, miR-21-5p was one of the few miRNAs, together with miR-223 and miR-155, which are directly related to osteoclast differentiation [45]. In an explorative analysis of 175 serum miRNAs in patients with recent OFs

and age-matched control, the down-regulation of miR-328-3p and let-7g-5p in response to OF were identified (Fold Change (FC)=-1.54, P=0.034 and FC=-1.24, P=0.041, respectively) and later confirmed (FC=-2.04, P=0.016 and FC=-1.89, P=0.027, respectively) in an independent case-control sample[46].

Furthermore in 2016, Kocijan R *et al.* designed a case-control study including a cross-sectional design of 36 patients with prevalent low-traumatic fractures and 39 control subjects, one hundred eighty-seven miRNAs were quantified in serum by qPCR, compared between groups and correlated with established bone turnover markers. Eight serum miRNAs (miR-152-3p, miR-30e-5p, miR-140-5p, miR-324-3p, miR-19b-3p, miR-335-5p, miR-19a-3p, miR-550a-3p) were found down-regulated with low traumatic fractures, which excellently discriminated OF cases and controls, regardless of age and sex (AUC>0.9, P<0.05) [47]. In this way, miRNAs might be directly linked to bone tissue homeostasis because of the significant correlations between miR-29b-3p and P1NP[48], miR-365-5p and iPTH, TRAP5b, P1NP and Osteocalcin[49].

Mandourah AY*et al.* reported that the circulating levels of miR-4516 was significantly lower (P<0.001) in the OFs with low BMD than non-fracture subjects with low BMD or non-osteoporotic controls [50]. Significantly, Ramírez-Salazar EG *et al.* reported that serum miR-140-3p and miR-23b-3p, which were significantly elevated in OF vs. NF subjects, presented high performances in relation to OFs (AUC=0.92, P<0.0001; AUC=0.88, P<0.0001)[51].

To conclude, each miRNA has specific characteristics predicting risk of osteoporosis. For example, miR-214 inhibits bone formation via ATF4 and is down-regulated in osteoporotic bone. As concluded by

Chen et al. [52] there are biological differential expressions in some c-miRNAs between osteoporotic and non-osteoporotic individuals, but their study did not determine that these specific circulating miRNAs were moleculars of osteoporosis.

3. Potential metabolite moleculars

Metabolites, such as urinary mineral products, amino acid and lipid mediators, all play essential roles in bone metabolism as described above. Potential diagnostic metabolite moleculars of OFs were summarized and listed in **Table 5** and presented in the following three categories.

3.1 Urinary mineral moleculars

Elevated fasting urinary calcium/creatinine ratio mainly manifested the process of osteoporosis. The percentage of hypercalciuria in urine in OF women (40%) was significantly higher compared to non-OF women (18.8%) (*P*=0.04) in a Spanish cross-sectional study, as reported by Ochoa-Hortal Rull MA[53]. 24-hour urinary calcium and fasting calcium/creatinine ratio may show some great characters as potential diagnostic moleculars for OFs, to some extent [54]. However, these results were only obtained from epidemical studies, more researches demonstrated these values of clinical application are warrented.

3.2 Amino acid moleculars

Homocysteine (Hcy) level in serum or plasma was a potential diagnostic molecular of OFs. Kuroda T *et al.* discovered that the plasma tHcy level was a significant risk factor for severe VFs in postmenopausal Japanese women [OR=1.22(1.03-1.46)-1.27(1.04-1.58), P<0.05) [55], and the plasma tHcy level was significantly higher in moderate VFs (Grade 1, 9.2 ± 2.8 µM) and severe VFs (Grade 2, 9.8 ± 3.7 µM), as compared to the Grade 0 (8.9±2.9 µM) [55].Besides, plasma Hcy concentration differed significantly between OF group (17.21±5.99 µM), high-energy fracture (HEF) group (10.82±3.60µM) and non-bone-fracture (NF) group (12.91±4.32µM) (F=9.483), and was positively correlated with bone resorption markers (serum C-terminal cross linking telopeptide of type I collagen (S-CTX), plasma levels of 25-OH Vit D) related to OF fracture in elderly Chinese cohort[56]. Consistently, Bahtiri E *et al.* reported that serum Hcy level was inversely associated with both lumbar spine BMD (r=-0.163, P<0.05) and femur neck BMD (r=-0.164, P<0.05), supporting it as a risk factor of OFs [57]. Of note, reducing plasma Hcy level can improve bone quality and reduce the risk of falling in elderly patients secondary to bone or skeletal muscle weakness, thereby reducing the risk of OPFs in the elderly. But how to predict the occurrence of OPFs is the direction for future research.

3.3 Lipid mediators

Lipid mediators, such as S1P and retinol, also play an important role in diagnosing OFs. For instance, plasma S1P levels were markedly higher in subjects with VFs ($7.49\pm3.44\mu$ M) than in those without VFs ($5.58\pm2.01\mu$ M) [OR=9.33(2.68-32.49), *P*=0.001], and increased with the number of VFs in a dose-response manner (*P*_{for the trend} <0.001) [58]. An independent study reported that the adjusted OR (95%Cl) of plasma S1P levels for OFs was 2.05(1.03-4.00)[59].

Navarro-Valverde Cet al. reported that the serum retinol level was a risk factor of OFs, which was negatively correlated with BMD at lumbar spine (r=-0.210; P<0.01) and femoral neck (r=-0.324, P<0.001) after multivariable adjustment [60]. A particularly interesting finding is that the plasma S1P level was associated with the risk of incident fracture, even after additional adjustment for antiosteoporotic medication. These results could be partially explained by the potential effects of S1P on the response to the antiosteoporotic medication. In conclusion, plasma S1P level, in combination with the BMD, could be a useful marker for predicting the risk of osteoporotic fracture.

Conclusions and Future Prospective

OFs are associated with high mortality and morbidity, and seriously affect the life quality of patients worldwide. In this study, we systematically reviewed the international research progress on the other types of predictive and diagnostic moleculars for OFs, and summarized significant moleculars with potential clinical utility. Collectively, remarkable variations in protein, miRNA and metabolite levels with the risk of OFs were principally identified through case-control studies and/or longitudinal cohort studies in various ethnicities.

The achievements in OF molecular development, as reviewed above, from clinical observations or discoveries to the recent fundamental researches of bone biology, represent private and public attention, investments, and efforts towards understanding the pathogenesis and preventing the occurrence of OFs. As reported in many clinical trials, some protein moleculars, including IGFBP-1, periostin, adiponectin, CRP, BAP and FGF23, etc. are positively associated with incident OFs. Other proteins, such as IGF-1, IGFBP-3 and periostin, demonstrate great diagnostic values for OFs. Besides these significant protein moleculars, metabolite products, such as urinary mineral metabolite, amino acid and lipid mediators might be crucial markers, as well. Furthermore, we have also summarized some reliable miRNAs markers, which demonstrate great diagnostic values (AUC>0.9, P<0.05) in OFs.

In conclusion, great progress has been made in OF moleculars discovery in the past decade, and some novel and promising moleculars have emerged. To be noted, clinical data on these novel moleculars remain limited, and sometimes the epidemical results were controversial among independent study samples. For example, the miR-22-3p was found to be moderately up-regulated in the discovery cohort, but significant down-regulation was observed in the validation cohort[46].

Therefore, it is necessary and would be important to confirm the potential clinical utility of the novel moleculars through independent studies and with larger sample sizes. With global aging population increasing, senile osteoporosis and the resultant fractures incur heavy social and economic burden. The new discoveries with no doubt will greatly promote the development of reliable moleculars for OFs and inform development of novel tools and strategies for diagnosing, predicting, and preventing OP and OFs.

Conflict of interest statement

No conflict of interest were reported by all authors.

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References

- Briggs, A.M., et al., Musculoskeletal Health Conditions Represent a Global Threat to Healthy Aging: A Report for the 2015 World Health Organization World Report on Ageing and Health. Gerontologist, 2016. 56 Suppl 2: p. S243-55.
- Hinz L, F.E., Kline G, How Good is Our Best Guess? Clinical Application of the WHO FRAX Tool in Osteoporotic Fracture Risk Determination and Treatment Decisions. Calcif Tissue Int., 2016. 99(2): p. 114-20.
- 3. Liu SY, H.M., Chen R, et al., Comparison of strategies for setting intervention thresholds for Chinese postmenopausal women using the FRAX model. Endocrine., 2019.
- 4. Su Y, L.J., Kwok T., The role of previous falls in major osteoporotic fracture prediction in conjunction with FRAX in older Chinese men and women: the Mr. OS and Ms. OS cohort study in Hong Kong. Osteoporos Int., 2018. **29(2)**: p. 355-363.
- 5. Estrada, K., et al., *Genome-wide meta-analysis identifies 56 bone mineral density loci and reveals 14 loci associated with risk of fracture.* Nature Genetics, 2012. **44**(5): p. 491-501.
- Kemp, J.P., et al., Identification of 153 new loci associated with heel bone mineral density and functional involvement of GPC6 in osteoporosis. Nat Genet, 2017. 49(10): p. 1468-1475.
- Cauley, J.A., Osteoporosis: fracture epidemiology update 2016. Curr Opin Rheumatol, 2017.
 29(2): p. 150-156.
- Liu X, X.X., MicroRNA-137 dysregulation predisposes to osteoporotic fracture by impeding ALP activity and expression via suppression of leucine-rich repeat-containing G-protein-coupled receptor 4 expression. Int J Mol Med., 2018. 42(2): p. 1026-1033.
- Lundin H, S.M., Strender LE, et al., High Serum Insulin-Like Growth Factor-Binding Protein 1 (IGFBP-1) is Associated with High Fracture Risk Independent of Insulin-Like Growth Factor 1 (IGF-I). Calcif Tissue Int., 2016. 99(4): p. 333-9.
- Rousseau, J.C., et al., Serum periostin is associated with fracture risk in postmenopausal women: a 7-year prospective analysis of the OFELY study. J Clin Endocrinol Metab, 2014.
 99(7): p. 2533-9.
- Bonnet, N., et al., Serum Levels of a Cathepsin-K Generated Periostin Fragment Predict Incident Low-Trauma Fractures in Postmenopausal Women Independently of BMD and FRAX.
 J Bone Miner Res, 2017. 32(11): p. 2232-2238.
- 12. Johansson, H., et al., High serum adiponectin predicts incident fractures in elderly men:

Osteoporotic fractures in men (MrOS) Sweden. J Bone Miner Res, 2012. 27(6): p. 1390-6.

- Johansson, H., et al., Waning predictive value of serum adiponectin for fracture risk in elderly men: MrOS Sweden. Osteoporos Int, 2014. 25(7): p. 1831-6.
- 14. Oei, L., et al., Dissecting the relationship between high-sensitivity serum C-reactive protein and increased fracture risk: the Rotterdam Study. Osteoporos Int, 2014. **25**(4): p. 1247-54.
- Tamaki, J., et al., Biochemical markers for bone turnover predict risk of vertebral fractures in postmenopausal women over 10 years: the Japanese Population-based Osteoporosis (JPOS) Cohort Study. Osteoporos Int, 2013. 24(3): p. 887-97.
- Mirza, M.A., et al., Serum fibroblast growth factor-23 (FGF-23) and fracture risk in elderly men. J Bone Miner Res, 2011. 26(4): p. 857-64.
- Lane, N.E., et al., Association of serum fibroblast growth factor 23 (FGF23) and incident fractures in older men: the Osteoporotic Fractures in Men (MrOS) study. J Bone Miner Res, 2013. 28(11): p. 2325-32.
- Gossiel, F., et al., Effect of Teriparatide Treatment on Circulating Periostin and Its Relationship to Regulators of Bone Formation and BMD in Postmenopausal Women With Osteoporosis. J Clin Endocrinol Metab, 2018. 103(4): p. 1302-1309.
- 19. Szulc, P., et al., Lower fracture risk in older men with higher sclerostin concentration: a prospective analysis from the MINOS study. J Bone Miner Res, 2013. **28**(4): p. 855-64.
- 20. Tamaki J, K.K., Fujita Y, Iki M, Yura A, Miura M, Sato Y, Okamoto N, Kurumatani N., *Ratio of Endogenous Secretory Receptor for Advanced Glycation End Products to Pentosidine Predicts Fractures in Men.* J Clin Endocrinol Metab., 2018. **103(1)**: p. 85-94.
- Garnero P, H.E., Chapuy MC, Marcelli C, Grandjean H, Muller C, Cormier C, Bréart G, Meunier
 PJ, Delmas PD, Markers of bone resorption predict hip fracture in elderly women: the EPIDOS
 Prospective Study. J Bone Miner Res., 1996. 11(10): p. 1531-8.
- 22. Garnero P, C.P., Sornay-Rendu E, Qvist P, Delmas PD., Type I collagen racemization and isomerization and the risk of fracture in postmenopausal women: the OFELY prospective study. J Bone Miner Res., 2002. **17(5)**: p. 826-33.
- Yoshimura, N., et al., Biochemical markers of bone turnover as predictors of osteoporosis and osteoporotic fractures in men and women: 10-year follow-up of the Taiji cohort. Mod Rheumatol, 2011. 21(6): p. 608-20.
- 24. Krege JH, L.N., Harris JM, Miller PD., PINP as a biological response marker during teriparatide

treatment for osteoporosis. Osteoporos Int., 2014. 25(9): p. 2159-71.

- Meesters, D.M., et al., Malnutrition and Fracture Healing: Are Specific Deficiencies in Amino Acids Important in Nonunion Development? Nutrients, 2018. 10(11).
- 26. Gjesdal, C.G., et al., *Plasma homocysteine, folate, and vitamin B 12 and the risk of hip fracture: the hordaland homocysteine study.* J Bone Miner Res, 2007. **22**(5): p. 747-56.
- 27. Enneman, A.W., et al., *The association between plasma homocysteine levels, methylation capacity and incident osteoporotic fractures.* Bone, 2012. **50**(6): p. 1401-5.
- 28. Oyen, J., et al., Plasma dimethylglycine, nicotine exposure and risk of low bone mineral density and hip fracture: the Hordaland Health Study. Osteoporos Int, 2015. **26**(5): p. 1573-83.
- 29. Cabrera, D., et al., Association of Plasma Lipids and Polar Metabolites with Low Bone Mineral Density in Singaporean-Chinese Menopausal Women: A Pilot Study. Int J Environ Res Public Health, 2018. **15**(5).
- Lecka-Czernik B, B.S., Stechschulte LA, Chougule AS, Marrow Fat-a New Target to Treat Bone Diseases? Curr Osteoporos Rep., 2018. 16(2): p. 123-129.
- Ahn, S.H., et al., Association of Bone Marrow Sphingosine 1-phosphate Levels with Osteoporotic Hip Fractures. J Bone Metab, 2013. 20(2): p. 61-5.
- Ardawi, M.M., et al., High Plasma Sphingosine 1-phosphate Levels Predict Osteoporotic Fractures in Postmenopausal Women: The Center of Excellence for Osteoporosis Research Study. J Bone Metab, 2018. 25(2): p. 87-98.
- Ginsberg, C., et al., The 24,25 to 25-hydroxyvitamin D ratio and fracture risk in older adults: The cardiovascular health study. Bone, 2018. 107: p. 124-130.
- Swanson, C.M., et al., Associations of 25-Hydroxyvitamin D and 1,25-Dihydroxyvitamin D
 With Bone Mineral Density, Bone Mineral Density Change, and Incident Nonvertebral
 Fracture. J Bone Miner Res, 2015. 30(8): p. 1403-13.
- 35. Cauley, J.A., et al., Serum 25-hydroxyvitamin D and the risk of hip and nonspine fractures in older men. J Bone Miner Res, 2010. **25**(3): p. 545-53.
- 36. Robinson-Cohen, C., et al., *Mineral metabolism markers and the long-term risk of hip fracture: the cardiovascular health study.* J Clin Endocrinol Metab, 2011. **96**(7): p. 2186-93.
- 37. Harris, T.B., et al., Plasma phospholipid fatty acids and fish-oil consumption in relation to

osteoporotic fracture risk in older adults: the Age, Gene/Environment Susceptibility Study. Am J Clin Nutr, 2015. **101**(5): p. 947-55.

- 38. Sugimoto T, N.K., Kuribayashi F, Chihara K,et al., Serum Levels of Insulin-like Growth Factor (IGF) I, IGF-Binding Protein (IGFBP)-2, and IGFBP-3 in Osteoporotic Patients with and without Spinal Fractures. JOURNAL OF BONE AND MINERAL RESEARCH, 1997. **12(8)**: p. 1272-9.
- Nakaoka D, S.T., Kaji H, Kanzawa M, Yano S, et al., Determinants of bone mineral density and spinal fracture risk in postmenopausal Japanese women. Osteoporos Int., 2001. 12(7): p. 548-54.
- Yamaguchi, T., et al., Serum levels of insulin-like growth factor (IGF); IGF-binding proteins-3,
 -4, and -5; and their relationships to bone mineral density and the risk of vertebral fractures in postmenopausal women. Calcif Tissue Int, 2006. 78(1): p. 18-24.
- 41. Kim, B.J., et al., Plasma periostin associates significantly with non-vertebral but not vertebral fractures in postmenopausal women: Clinical evidence for the different effects of periostin depending on the skeletal site. Bone, 2015. **81**: p. 435-441.
- Yan, J., et al., Circulating periostin levels increase in association with bone density loss and healing progression during the early phase of hip fracture in Chinese older women. Osteoporos Int, 2017. 28(8): p. 2335-2341.
- 43. Seeliger C, K.K., Haug AT, Vester H, Schmitt A, Bauer JS, van Griensven M, Five freely circulating miRNAs and bone tissue miRNAs are associated with osteoporotic fractures. J Bone Miner Res, 2014. **29(8)**: p. 1718-28.
- Panach, L., et al., Serum Circulating MicroRNAs as moleculars of Osteoporotic Fracture. Calcif Tissue Int, 2015. 97(5): p. 495-505.
- Lian, J.B., et al., *MicroRNA control of bone formation and homeostasis*. Nat Rev Endocrinol, 2012. 8(4): p. 212-27.
- 46. Weilner, S., et al., *Differentially circulating miRNAs after recent osteoporotic fractures can influence osteogenic differentiation.* Bone, 2015. **79**: p. 43-51.
- Kocijan, R., et al., Circulating microRNA Signatures in Patients With Idiopathic and Postmenopausal Osteoporosis and Fragility Fractures. J Clin Endocrinol Metab, 2016. 101(11): p. 4125-4134.
- 48. Li Z, H.M., Jafferji M, et al., *Biological functions of miR-29b contribute to positive regulation* of osteoblast differentiation. J Biol Chem. 2009. **284(23)**: p. 15676–15684.

- 49. Franceschetti T, D.N., Kessler CB, et al., *Pathway analysis of microRNA expression profile during murine osteoclastogenesis*. . PLoS One . 2014. **9(9)**: p. e107262.
- 50. Mandourah, A.Y., et al., *Circulating microRNAs as potential diagnostic moleculars for osteoporosis.* Sci Rep, 2018. **8**(1): p. 8421.
- 51. Ramirez-Salazar, E.G., et al., Serum miRNAs miR-140-3p and miR-23b-3p as potential moleculars for osteoporosis and osteoporotic fracture in postmenopausal Mexican-Mestizo women. Gene, 2018. **679**: p. 19-27.
- 52. Chen, Z., M.G. Bemben, and D.A. Bemben, *Bone and muscle specific circulating microRNAs in postmenopausal women based on osteoporosis and sarcopenia status.* Bone, 2019. **120**: p. 271-278.
- 53. Rull, M.A., et al., *The importance of urinary calcium in postmenopausal women with osteoporotic fracture.* Can Urol Assoc J, 2015. **9**(3-4): p. E183-6.
- 54. Wu, V.C., et al., *Risk of Fracture in Primary Aldosteronism: A Population-Based Cohort Study.* J Bone Miner Res, 2017. **32**(4): p. 743-752.
- 55. Kuroda, T., et al., *Plasma level of homocysteine associated with severe vertebral fracture in postmenopausal women.* Calcif Tissue Int, 2013. **93**(3): p. 269-75.
- 56. Zhu, Y., et al., *Plasma homocysteine level is a risk factor for osteoporotic fractures in elderly patients*. Clin Interv Aging, 2016. **11**: p. 1117-21.
- 57. Bahtiri E, I.H., Rexhepi S, Qorraj-Bytyqi H, Thaçi K, et al., *Relationship of homocysteine levels with lumbar spine and femur neck BMD in postmenopausal women.* Acta Reumatol Port., 2015. **40(4)**: p. 355-62.
- 58. Kim, B.J., et al., *Plasma sphingosine 1-phosphate levels and the risk of vertebral fracture in postmenopausal women.* J Clin Endocrinol Metab, 2012. **97**(10): p. 3807-14.
- Bae, S.J., et al., The circulating sphingosine-1-phosphate level predicts incident fracture in postmenopausal women: a 3.5-year follow-up observation study. Osteoporos Int, 2016. 27(8):
 p. 2533-41.
- Navarro-Valverde, C., et al., High Serum Retinol as a Relevant Contributor to Low Bone Mineral Density in Postmenopausal Osteoporotic Women. Calcif Tissue Int, 2018. 102(6): p. 651-656.

Biomarker	Reference	Study Design	Sample	Sample Size (N=OF+NF or N)	Gender	Ethnicity /Population	Age(Mean ± S.D. or Mean) (years)	Follow-up (years)	Phenotype	Concentration	Effect Size	e or Difference
										https://	Male	Female
Insulin-like growth									by h	://do		HF: Adj.HR=1.46(1.08-1.99) pe
factor-binding protein 1	Lundin H et	Prospective	Serum	N-351	Female	Sweden	73(69-79)	10	OF =	N A	ΝΑ	SD increase.
(IGFBP-I)	al.2016[9]	Trospective	berum	11-301	Tennale	Sweden	15(0) 17)	10		0.1	1 1.1 1.	MOF: Adj.HR=1.33(1.05-1.69,
(101211)									ade a	101/		P=0.017)
Periostin	Rousseau JC et	Prospective	Serum	N=607	Female	French	66.6 + 8.4	7	VF lie	1249±340 ng/ml vs.	N.A.	RR=2.81(1.1-7.3),P=0.03
	al.2014[10]	1105peente	Seram	11 007	1 0111110				adura	0 1162±298 ng/ml		Adj.RR=1.88(1.1-3.2)
Periostin fragment	Bonnet N et							Mean	Low-traum	$^{22}_{20}$ 57.5 \square + \square 36.6 \square ng/mL vs.		
resulting from cathepsin K	al.2017 [11]	Prospective	Serum	N=695	Female	Geneva	$65.0\Box \pm \Box 1.5$	4.7□±□1.9		$42.5\pm$ 23.4 mL	N.A.	HR=2.14(1.54-2.97), P<0.001
digestion (K-Postn)									fracture	129.tt		
Adiponectin	Johansson H et	Prospective	Serum	N=999	Male	Swedish	N.A.	Mean 5.2	OF OF	ອີ. 14.2± 8.9 μg/ml vs.	HR/SD = 1.46(1.23 - 1.72)	N.A.
<u>r</u>	al.2012[12]		~							^ω . 11.5±5.8 μg/ml	111000 1110(1120 1112)	
									4.0 I	n 90	Adj. for age and time since	
									nterr	sted	baseline HR=1.47(1.23-1.77)	
Adiponectin	Johansson H et	Prospective	Prospective Serum	N=989	Male	Swedish	70-81	Mean 5.3		4.4±9.3 μg/ml vs. 11.5±5.8	per SD	N.A.
	al.2014[13]							(0.0-7.4)	a lice	a σ μg/ml	Adj. for femoral neck BMD	
									cens	020.	and age HR=1.40(1.16-1.68)	
									e. c	The	per SD	
Serum high-sensitivity	Oei L et	- ·	a							copy	HR=1.0	6(1.02-1.11)
C-reactive protein (CRP)	al.2014[14]	Prospective	Serum	N=6386	59% Female	Rotterdam	69.1	Mean 11.6	OF S	N.A.	HF:HR=1	.09(1.02-1.17)
							547 (NOM -5		bio	hold	VF:OR=1	.34(1.14-1.38)
Dono sposifio allealing	Tamaki Lat						54.7 (YSM<5			der fo		1.45, 12, 21
phosphatase (BAP)	al 2013[15]	Prospective	Serum	N=522	Female	Japanese	years); $66.6 (\text{VSM} > -5)$	Median 10	VF c	t N.A.	N.A.	(1.43-15.21)
phosphatase (BAI)	al.2013[13]								ipen	spre		$(1 \ 12 \ 174)$
							years)		ury.	print	OF HR = 1.20(1.03 - 1.40)	(1.12-1.77)
											VF:HR=1.33(1.02-1.75)	
Fibroblast growth factor 23	Mirza MA et	Prospective	Serum	N=2868	Male	Swedish	75.4□±□3.2	Median	OF/VF	N.A.	Above 55.7pg/mL:	N.A.
(FGF23)	al.2011[16]						3.35			HF:HR=2.30(1.16-4.58)		
											non-VF:HR=1.63(1.01-2.63)	
							*Q1:73.40±5.8				DH 0.00/1.07.0.70	
Fibroblast growth factor 23	Lane NE et	Due of the	6 - ·	N 1600	MI	C 1' 1	*Q2:73.63±5.91	5.2	Nama' OF	N.A. (Q1-Q3: 4.2-22.4	KH=2.02(1.07-3.79) in men	NT A
(FGF23)	al.2013[17]	Prospective	Serum	N=1680	Male	Swedish	*Q3:74.40±6.0	5.3	Nonspine OF	pg/mi; vs. Q4: 22.4-111.1	with eur Kuruys	N.A.
							*Q4:73.70±5.8			pg/mi)	<00 mL/mm/1./3 m2	

Sclerostin	Szulc P et al.2013[19]	Prospective	Serum	N=710	Male	French	65±7	10	OF	0.61 (0.49, 0.77)ng/mL	All F:HR=0.73(0.58-0.92)per SD, P<0.05 MOF:HR=0.67(0.50-0.91)per SD, P<0.01	N.A.
PEN	Tamaki J et al.2018[20]	Prospective	Serum	N=1285	Male	Japanese	73.1□±□5.1	5	OF	0.057 vs. 0.049 μg/ml	FN:HR=1.48(1.00-2.18) LS:HR=1.51(1.03-2.21)	N.A.
esRAGE-to-PEN ratio	Tamaki J et al.2018[20]	Prospective	Serum	N=1285	Male	Japanese	73.1□±□5.1	5	OF =	3.6 ± 1.8 vs. 4.9 ± 1.9 (geometric mean \pm SD)	FN:HR=0.67(0.45-0.98) LS:HR=0.64 (0.43-0.95)	N.A.
C-telopeptide (CTX)	Garnero P et al.1996[21]	Prospective	Serum	N=7598	Female	French	>=75	Mean 22 months	HF availa	266 ±150μg/mmol Cr vs.	N.A.	OR=2.2(1.3-3.6)
C-telopeptide (CTX)	Garnero P et al.2002[22]	Prospective	Serum	N= 408	Female	French	64(50-89)	Median 6.8	VF and b peripheral fractures	22.8±8.0 μmmol/mmol Cr 22.8±8.0 μmmol/mmol Cr 2500 km s. 21.2±8.5 μmmol/mmol	N.A.	RR=4.5(2.0-10.1)
C-terminal cross-linking telopeptide of type I collagen (beta-CTX)	Yoshimura N et al.2011[23]	Prospective	Serum	N=400	Male/female =1/1	Japanese	Male:59.4 Female:59.8	10	Spinal OF	Male:0.187 vs. 0.121 ng/ml Female:0.234 vs. 0.145 res ng/ml	N.S.	HR=1.80, P<0.001
N-terminal cross-linking telopeptide of type I collagen (NTX)	Yoshimura N et al.2011[23]	Prospective	Serum	N=400	Male/female =1/1	Japanese	Male:59.4 Female:59.8	10	4.0 Internation	Male:13.6 vs. 4.2 nmolBCE/L Female:15.2 vs. 4.0 nmolBCE/L	N.S.	HR=1.96, P<0.01
N-terminal propeptide of type I procollagen (PINP)	Yoshimura N et al.2011[23]	Prospective	Serum	N=400	Male/female =1/1	Japanese	Male:59.4 Female:59.8	10	Spinal OF	5°_{0} Male: 38.2 vs. 19.5 ng/ml	HR=2.80, <i>P</i> < 0.05	HR=1.65, <i>P</i> <0.05

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Biomarker	Reference	Study Design	Sample	Sample Size (N=OF+NF or N)	Gender	Ethnicity /Population	Age(Mean ± S.D. or Mean) (years)	Follow-up (years)	ot certification of the state o	Concentration	Effect Size or Difference	
									doi.org / peer It		Male	Female
free deoxypyridinoline	Garnero P et al.1996[21]	Prospective	Urine	N=7598	Female	French	>=75	Mean 22 months	10.1101/2020.03 review) Mthe aut is made available	6.41±2.20nmol/mmol Cr vs. 5.80±1.94nmol/mmol Cr	N.A.	OR=1.9(1.1-3.2)
urinary deoxypyridinoline	Yoshimura N et al.2011[23]	Prospective	Urine	N=400	Male/female =1/1	Japanese	Male:59.4 Female:59.8	10	.31.2004@29.this hor/fundeat who ha under a @C-BY-N S	Male:3.06 vs. 1.53 pmol/lmolCr Female:4.76 vs. 1.91 pmol/lmolCr	N.S.	HR=1.40, P<0.05
homocysteine	Gjesdal CG et al.2007[26]	Prospective	Plasma	N=4766	Male/female =2127/2639	Bergen	65-67 years at enrollment	Median 12.6	version post s gra fie d m C-ND母.0 Int	7–20 µM	N.S.	HR=2.42(1.43-4.09)
homocysteine	Enneman AW et al.2012[27]	Prospective	Plasma	N=503	Female	Rotterdam	68.5(61.3–74.9)	Mean 7	ed April 6, 2 edRate a lic ernational I	(9.4–11.6)µmol/L	N.A.	Adj. for age, BMI, FN-BMD. HR=1.82(1.02-3.22)
Dimethylglycine(DMG)	Øyen J et al.2015[28]	Longitudinal	Plasma	N=3310	Male/female =1017/1204	Bergen, Norway	71-74	Mean 10.8	enster .	(4.4±1.5)µmol/L	Adj. for BM	/ID, HR=1.70(1.28-2.26)
Sphingosine-1-phosphate (S1P)	Ardawi MM et al.2018[32]	Prospective longitudinal	Plasma	N=707	Female	Saudi Arabia	61(53-93)	Mean (5.2±1.3)	e copyright display the	7.23±0.79 μmol/L vs.5.02±0.51 μmol/L	N.A.	Adj.HR=6.12(4.92-7.66) per SD increase.
24,25-dihydroxyvitmin D [24,25(OH)2D]	Ginsberg C et al.2018[33]	Longitudinal	Serum	N=890	Male/female =267/178	16% African-American	78	Mean 8.4	∶ holder for pre∰int ir	(1.7±1.0) ng/ml	HR=0.73(0	61-0.87) per SD increase
the ratio of 24,25(OH)2D to 25(OH)D [VMR]	Ginsberg C et al.2018[33]	Longitudinal	Serum	N=890	Male/female = 267/178	16% African-American	78	Mean 8.4	n pertectui	(6.84±2.23) ng/ml	HR=0.74(0	61-0.88) per SD increase
5-hydroxyvitaminD(250HD)	Swanson CM et al.2015[34]	Prospective	Serum	N=1000	Male	USA	74.6±6.2	Mean 5.1	옷 ਜੂਂ non-VF	3.13 vs. 20.90 ng/mL	HR=2.01(1.24-3.26)	N.A.
1,25-dihydroxyvitaminD (1,25(OH)2D)	Swanson CM et al.2015[34]	Prospective	Serum	N=1000	Male	USA	74.6±6.2	Mean 5.1	non-VF	8.70 vs. 51.60 pg/mL	HR=1.99(1.19-3.33)	N.A.
Polyunsaturated fatty acids (PUFAs)	Harris TB et al.2015[37]	Longitudinal	Plasma	N=540+898	OF:male/fe male = 19/35 NF: male/female	N.A.	OF:78.5± 5.69 NF:76.5± 5.48	Mean 7	OF	N.A.	HR=0.60(0.41-0.89)	Inverse association, P-trend = 0.0ϵ

Notes: OF: osteoporotic fracture; NF: non-fracture; VF: vertebral fracture; HF: hip fracture; FN: femoral neck; HR: hazard ratio; OR: odds ratio.

*N.A.: not available; N.S. not significant.

Biomarker	Reference	Study Design	Sample	Sample Size (N=OF+NF or N)	Gender	Ethnicity/ Population	Age(Mean ± S.D. or Mean) (years)	Phenotype	Concentration	Effect Size or Difference
IGF-I	Sugimoto T et al.1997[38]	Cross-sectional	Serum	N=165	Female	Japanese	62	Spinal OF	os://doi.org/10	Lower in OF, P<0.01; Cutoff 110 ng/ml provided a specificity of 81% with a sensitivit of 76%.
IGF-I	Nakaoka D et al.2001[39]	Cross-sectional	Serum	N=205	Female	Japanese	64	Spinal OF _{avai} is Spinal OF _{avai}		OR=0.19(0.07-0.59), P=0.003
IGF-I	Yamaguchi T et al.2006[40]	Cross-sectional	Serum	N=61+132	Female	Japanese	62.5(46-88)	able under VF	297.1±32.1 ng/dL vs. 143.9±40.9 ng/dL	OR=0.29(0.15-0.57), P=0.0003
IGFBP-3	Sugimoto T et al.1997[38]	Cross-sectional	Serum	N=165	Female	Japanese	62	Spinal OF	1494229. ➡ N.A. ➡ is <	Lower in OF, P<0.01; Cutoff 2.1µg/ml provided a specificity of 81% with a sensitivit of 81%.
IGFBP-3	Yamaguchi T et al.2006[40]	Cross-sectional	Serum	N==61+132	Female	Japanese	62.5 (46-88)	VF 4.0 In	$\frac{g}{g}$ 2.18±1.02 µg/mL vs. 3.23±1.07 μ g/mL	OR=0.31(0.16-0.61), P=0.0007
Periostin	Kim BJ et al.2015[41]	Case-control	Plasma	N==133+133	Female	Korean	62.7±6.4 Cases; 63.0±6.3 Control	ternational license OF	³² 32.4 ng/mL vs. 29.2 ng/mL, any ⁵² fracture; ⁵² 32.6 ng/mL vs. 28.6 ng/mL, ⁵³ non-VF.	Any fracture: OR=1.50(1.14-1.97), P=0.003 Non-VF: OR=1.59(1.12-2.24), P=0.009
Periostin	Yan J et al.2017[42]	Case-control	Serum	N==261+106	Female	Chinese	80 (76–84) Cases; 78 (72–83) Controls	e. HF HF	ਸੋ 89 45.51 (28.77–79.70) ng/mL vs. ਤੋੜੋ 32.87 (19.71–53.58) ng/mL ਰੁ	FN-BMD: β(standardized coefficient)=–0.390,P=0.004 LS-BMD: β(standardized coefficient)=–0.158,P=0.190

Table 4: Diagno	stic microRNAs moleculars (of Osteoporotic F	`ractures		(which was		
microRNA ID	Reference	Study Design	Sample Size (N=OF+NF or N)	Sample	Expression Trend in Street Case vs. Control	Fracture Diagnostic Value or Difference	Phenotype
miR-21	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum and Bone tissue	Up-regulated	AUC:0.63(0.53–0.73;P= 0.013)	OF
miR-23a	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum and Bone tissue	Up-regulated w made w made	AUC:0.63(0.53–0.73;P=0.015)	OF
miR-24	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum and Bone tissue	Up-regulated e utro	AUC:0.63 (0.53–0.74;P=0.013)	OF
miR-93	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum	Up-regulated a CO	AUC:0.68 (0.58–0.78;P=0.001)	OF
miR-100	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum and Bone tissue	Up-regulated C-20	AUC:0.69(0.60-0.78;P=0.0003)	OF
miR122a	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum	Up-regulated ne	AUC:0.77(0.69–0.86,P<0.0001)	OF
miR□124a	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum	Up-regulated brill	AUC:0.69(0.59–0.78;P=0.0005)	OF
miR□125b	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum	Up-regulated . The	AUC:0.76(0.67–0.85;P< 0.0001)	OF
miR□148a	Claudine Seeliger et al.2014[43]	Case-control	Serum:N=30+30 Bone:N=20+20	Serum	Up-regulated	AUC:0.61 (0.51–0.72;P=0.038)	OF
miR-21-5p	Layla Panach et al.2015[44]	Case-control	N=15+12	Serum	Up-regulated	Fold change=1.57, P=0.002	OF
miR-122-5p	Layla Panach et al.2015[44]	Case-control	N=15+12	Serum	Up-regulated	Fold change=5.48, P=0.00008	OF
miR-125b-5p	Layla Panach et al.2015[44]	Case-control	N=15+12	Serum	주 <u>ㅋ</u> Up-regulated	Fold change=4.62, P=0.0003	OF
miR-328-3p	Sylvia Weilner et al.2015[46]	Cross-sectional	N=37	Serum	Down-regulated	Fold change=-1.54, -2.04; P<0.05	OF
let-7g-5p	Sylvia Weilner et al.2015[46]	Cross-sectional	N=37	Serum	Down-regulated	Fold change=-1.24, -1.89; P<0.05	OF
miR-19a-3p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulated	AUC:0.929 (0.856–0.983)	Low traumatic fractures
miR-19b-3p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulated	AUC:0.944 (0.879–0.997)	Low traumatic fractures

miR-30e-5p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulated	AUC:0.959 (0.901–0.997)	Low traumatic fractures
miR-140-5p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulated	AUC:0.947 (0.900-0.983)	Low traumatic fractures
miR-152-3p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulated	AUC:0.962 (0.918-0.993)	Low traumatic fractures
miR-324-3p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulatee	AUC:0.950 (0.885–0.994)	Low traumatic fractures
miR-335-5p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulate	AUC:0.939 (0.872–0.986)	Low traumatic fractures
miR-550a-3p	Kocijan R et al.2016[47]	Case-control	N=36+39	Serum	Down-regulated ra	AUC:0.909 (0.837–0.970)	Low traumatic fractures
hsa-miR-4516	Mandourah AY et al.2018[50]	Case-control	N=139	Serum and Plasma	Up-regulated by the second	P = 0.00014	OF
miR-140-3p	Ramírez-Salazar EG et al.2018[51]	Case-control	N=20+20	Serum	Up-regulated 0.4 resident	AUC:0.92;P< 0.0001	OF
miR-23b-3p	Ramírez-Salazar EG et al.2018[51]	Case-control	N=20+20	Serum	Up-regulated Ation	AUC:0.88;P< 0.0001	OF
Notes: OF: osteopo	protic fracture; NF: non-fracture; AUC	2: area under curve.			6, 2020. The copyright holder for this preprint a license to display the preprint in perpetuity. fal license .		

Biomarker	Reference	Study Design	Sample	Sample Size (N=OF+NF or N)	Gender	Ethnicity/ Population	Age(Mean ± S.D. or Mean) (years)	not certified by	Concentration	Effect Size	or Difference in Concentration or Percentage
								doi.org		Male	Female
urinary calcium	Ochoa-Hortal Rull MA et al.2015[53]	Cross-sectional	Urine	N=55+32	Female	Spanish	OF:70.1±13.8 non-OF: 56.7±6.4	is made av	9.19±0.38mg/dL vs. 8.95±0.73mg/dL	N.A.	40% vs 18.8% hypercalciuria, P=0.04
homocysteine	Kuroda T et al.2013[55]	Cross-sectional	Plasma	N=1475	Female	Japanese	66.6±9.0	F VF 020.03.31.20049- the author/funder vailable under a C	Grade0: 8.9±2.9 μM Grade1: 9.2 ± 2.8 μM Grade2: 9.8 ± 3.7 μM Grade3: 11.3 ± 5.3 μM	N.A.	OR=1.22(1.03-1.46)-1.27(1.04 -1.58), P<0.05
homocysteine	Zhu Y et al.2016[56]	Cross-sectional	Plasma	N=82	Male/female =14/27	Chinese	80.61±5.61 (OF:81.82±5.49; HEF:78.88±5.75; non-bone-fracture:79.75±5.47)	F F Also this version p who has granted C-BY-NC-ND 4.C	OPF:17.21±5.99 μM HEF:10.82±3.60 μM NF:12.91±4.32 μM		F=9.483, P= 0.000
homocysteine	Bahtiri E et al.2015[57]	Cross-sectional	Serum	N=139	Female	N.A.	56.53±5.84	osted April I medRxiv	16.84±11.71 μM vs. 13.09±2.53 μM	N.A.	LS-BMD(r=-0.163,P<0.05) FN-BMD (r=-0.164,P<0.05)
Sphingosine-1-pho sphate (S1P)	Kim BJ et al.2012[58]	Case-control	Plasma	N=69+69	Female	Korean	VF:65.1±6.7 non-VF:64.9±6.5	6, 2020. T a license t nal license	7.49±3.44 μM vs. 5.58±2.01 μM	N.A.	OR=9.33(2.68-32.49)
Sphingosine-1-pho sphate (S1P)	Bae SJ et al.2016[59]	Longitudinal	Plasma	N=248	Female	Korean	Untreated: 59.1±7.7 Treated:58.4±6.3	. he copyright holder o display the preprin	OF	highest tertile: (5.90–16.5 1)µmol/l	N.A.
Retinol	Navarro-Valverde C et al.2018[60]	Cross-sectional	Serum	N=154	Female	Spanish	> 65 years	F or this preprin t in perpetuity.	N.A.	N.A.	LS-BMD (r=-0.210;P<0.01) FN-BMD(r=-0.324,P<0.001)

Notes: OF/OPF: osteoporotic fracture; NF: non-fracture; VF: vertebral fracture; HF: hip fracture; HEF: high-energy fracture; LS: lumbar spine; FN: femoral neck; OR: odds ratio. *N.A.: not available; N.S. not significant.