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Evaluation of saline, phosphate buffered saline and minimum essential medium as potential alternatives to viral transport media for SARS-CoV-2 testing

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Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2), the causative agent of coronavirus disease-19 (COVID-19), has caused a global pandemic since being discovered in late 2019. In response, clinical microbiology and public health laboratories have worked to develop, validate, and implement molecular assays to detect SARS-CoV-2 from respiratory samples. The preferred and most commonly collected specimen is a nasopharyngeal (NP) swab placed in viral transport media (VTM). As testing demand has increased, specimen collection and transportation supplies, including VTM, are decreasing nationwide. Due to these shortages of collection supplies and transport media, we assessed the feasibility of placing NP swabs in sterile $0.9 \%$ saline (Baxter; Deerfield, IL), sterile phosphate buffered saline (PBS), or minimum essential media (MEM) (Corning; Corning, NY) prior to testing for SARS-CoV-2 by a commercially-available (emergency use authorized [EUA]) FDA platform (cobas® SARS-CoV2; Roche Diagnostics, Indianapolis, IN) and a SARS-CoV-2 laboratory-developed test (LDT) that has been validated and submitted to the Food and Drug Administration for EUA approval. The Roche cobas SARS-CoV-2 test is performed on the cobas® 6800 platform (Roche) per the manufacturer's protocol. The SARS-CoV-2 LDT is performed as described in the supplemental materials, targeting the nucleocapsid (NUC) and open reading frame (ORF) genes of the virus.

For this study, samples were prepared by placing analyte-negative NP swabs into twelve $15-\mathrm{mL}$ conical tubes (Corning) containing 3-mL of either M4-RT VTM (Remel Inc; San Diego, CA), MEM, saline, or PBS for a total of 48 samples. Subsequently, each sample was spiked with SARS-CoV-2 positive patient material at a concentration of 10,000 copies $/ \mathrm{mL}$, which is 2 -fold higher than the defined limit of detection of the LDT. Two $15-\mathrm{mL}$ conical tubes containing 3-mL of each media (i.e., 8 total samples) functioned as negative controls. On day 0 (i.e., the day the samples were prepared), 6 contrived samples in each of the four media types listed above (i.e., 24
samples), as well as negative controls, were tested by the Roche cobas and LDT SARS-CoV-2 methods (Table 1). Following initial testing, half of the contrived samples were stored refrigerated $\left(2-8^{\circ} \mathrm{C}\right)$, while the remaining aliquots were stored frozen $\left(-15\right.$ to $\left.-25^{\circ} \mathrm{C}\right)$. The aliquots were pulled from storage on days 1,3 , and 7 and tested by both methods. Equivalence (i.e., qualitative results as well as $+/-2$ cycle threshold $\left[\mathrm{C}_{\mathrm{t}}\right]$ values) and stability ( $+/-2 \mathrm{C}_{\mathrm{t}}$ values over 7 days) of the alternative transport media were compared to VTM.

The SARS-CoV-2 results of both assays showed equivalence (i.e., $100 \%$ qualitative agreement and $\mathrm{C}_{\mathrm{t}}$ variation < 2 cycles) when swabs were stored in MEM, PBS, saline and VTM over 7 days at both refrigerated and frozen storage conditions (Table 1). No evidence of loss in sensitivity or stability (>2 Ct value increase) was observed for any of the transport media. One sample (PBS-2) showed lower (i.e., more sensitive) $\mathrm{C}_{\mathrm{t}}$ values on days 1,3 , and 7 . This may indicate slight variation in preparing the contrived samples. Internal control results for all samples were within established QC ranges (data not shown). Negative controls were tested on Day 0 and produced expected results, demonstrating that the media were free of SARS-CoV-2 contamination. Positive and negative extraction/amplification controls run with each plate produced expected results (data not shown). These data support the use of MEM, PBS, or $0.9 \%$ saline as alternatives to VTM for SARS-CoV-2 testing.
 saline

$1 \quad N$, number of samples tested; LDT, laboratory-developed test; NUC, nucleocapsid target; ORF, open reading frame target; E, envelope
2 target; Ct, cycle threshold; MEM, minimum essential medium; PBS, phosphate buffered saline

