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Environmental cleaning is effective for the eradication of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in contaminated hospital rooms: A patient from the Diamond Princess cruise ship

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To the Editor- Doctors, nurses, and other medical staff are greatly concerned about nosocomial outbreaks of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Environmental contamination is a possible source of nosocomial transmission [1, 2]. However, it is unclear how effective environmental cleaning is against SARS-CoV-2.

A 75-year-old man infected with SARS-CoV-2 was diagnosed as having COVID-19 during the quarantine period on the Diamond Princess cruise ship. He was transferred directly to our hospital on February 11, 2020. He resided in patient Room A for 2 days, then was moved to Room B, where he stayed for 19 days. After cleaning the rooms thoroughly with disinfectant (Rely⁺On Virkon, LANXESS, or RUBYSTA in Japan), we tested a total of 15 areas that were in close contact with the patient and medical staff. Swabs were used to transfer five environmental samples from Room A and 10 samples from Room B to universal transport media (Copan, Murrieta, CA). Cleaning was conducted immediately after the patient left the rooms. Environmental sampling was conducted within 5 days and 30 min after the patient left Rooms A and B, respectively. Nucleic acids were extracted using MagMAX Viral/Pathogen Nucleic Acid Isolation Kit (ThermoFisher Scientific, Waltham, MA), and examined by real-time reverse transcription polymerase chain reaction (RT-PCR) targeting the nucleocapsid (*N*) gene of SARS-CoV-2. Seven sets of primers and probes (CDC-N1, CDC-N2, CDC-N3, YCH-N1, YCH-N2, NIID-N1, and NIID-N2) were used to detect SARS-CoV-2 as previously described (Supplemental Table 1) [3]. For the internal positive control, the human ribonuclease P 30 subunit (*RPP30*) gene was used. The patient's records, timing of cleaning and sampling, and RT-PCR results were collated.

On admission, the patient had fever (39 °C) and a mild cough (Supplemental Table 2). The chest X-ray and computed tomography scan on day 1 showed signs of pneumonia in both lungs. He received lopinavir/ritonavir and antibacterial therapy on day 2 but showed respiratory failure. He received supplemental oxygen from day 4 to 15. After careful clinical management, the patient's overall status improved. RT-PCR showed his sputum was positive for SARS-CoV-2 on day 11. Subsequently, nasopharyngeal swabs were negative on days 17, 22, and 29.

The patient stayed in Room A for 3 days, during which he had the SARS-CoV-2 infection. After cleaning Room A, five environmental samples were examined by RT-PCR. All samples were negative for SARS-CoV-2 and positive or negative for *RPP30* (Table 1).

After the patient left Room A, he resided in Room B for 20 days. Ten environmental samples were collected after cleaning. All ten samples from Room B were negative for SARS-CoV-2 and positive or negative for *RPP30* (Table 1).

SARS-CoV-2 is detectable in several types of clinical sample including Bronchial lavage fluid, nasopharyngeal swab, pharyngeal swab, sputum, saliva and stool [4, 5]. Transmission of SARS-CoV-2 via surfaces in hospitals is of great concern to medical staff and patients. Blocking the potential routes of transmission is essential for preventing the spread of SARS-CoV-2 [6]. A recent

study showed that environmental contamination can occur via contact with patients with SARS-CoV-2 and upper respiratory tract symptoms [7]. We found that, after cleaning, all areas were negative for SARS-CoV-2; therefore, thorough cleaning is sufficient for SARS-CoV-2 decontamination.

This study had several limitations. First, RT-PCR was not performed before cleaning because of the risk of nosocomial transmission. Therefore, a comparison of the viral loads of high-touch areas before and after cleaning is required. Second, this study involved a single patient, and further studies are required to confirm the findings.

In summary, our data indicate the effectiveness of environmental cleaning for SARS-CoV-2 decontamination. This information is useful for infection control strategies and may alleviate the concerns of medical staff.

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Conflict of Interest. None.

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Table 1. Real-time PCR analysis of environmental samples

Location	RT-PCR (number of samples)	
Patient Room A	SARS-CoV-2	Human <i>RPP30</i>
Light switch	Negative (0/1)	Negative (0/1)
Nurse call attached to the bed	Negative (0/1)	Positive (1/1)
Toilet door handle	Negative (0/1)	Negative (0/1)
Bed guard	Negative (0/1)	Positive (1/1)
Anterior Room A		
Dust box	Negative (0/1)	Positive (1/1)
Patient Room B		
Bed desk	Negative (0/1)	Negative (0/1)
Bed guard	Negative (0/1)	Negative (0/1)
Door handle	Negative (0/1)	Positive (1/1)
Dust box, room side	Negative (0/1)	Negative (0/1)
Dust box, corridor	Negative (0/1)	Positive (1/1)
Control panel on mechanical ventilation	Negative (0/1)	Positive (1/1)
Light switch	Negative (0/1)	Negative (0/1)
Nurse call	Negative (0/1)	Positive (1/1)
Hand soap dispenser	Negative (0/1)	Negative (0/1)
Anterior Room B		
Sink, external rim and internal bowl	Negative (0/1)	Positive (1/1)

RPP30, ribonuclease P 30 subunit; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2

Supplemental Material

Supplemental Table 1. Sequences of the primers and probes used in the CDC, NIID, and YCH assays

Institute	Primer name	Description	Oligonucleotide Sequence (5'>3')	Modification
CDC	CDC-N1-F	2019-nCoV_N1 Forward Primer	5'-GACCCCAAAATCAGCGAAAT-3'	None
CDC	CDC-N1-R	2019-nCoV_N1 Reverse Primer	5'-TCTGGTACTGCCAGTTGAATCTG-3'	None
CDC	CDC-N1-P	2019-nCoV_N1 Probe	5'-FAM-ACCCCGCATTACGTTTGGTGGACC-BHQ1-3'	FAM/BHQ1
CDC	CDC-N2-F	2019-nCoV_N2 Forward Primer	5'-TTACAAACATTGGCCGCAAA-3'	None
CDC	CDC-N2-R	2019-nCoV_N2 Reverse Primer	5'-GCGCGACATCCGAAGAA-3'	None
CDC	CDC-N2-P	2019-nCoV_N2 Probe	5'-FAM-ACAATTTGCCCCAGCGCTTCAG-BHQ1-3'	FAM/BHQ1
CDC	CDC-N3-F	2019-nCoV_N3 Forward Primer	5'-GGGAGCCTTGAATACCAAAA-3'	None
CDC	CDC-N3-R	2019-nCoV_N3 Reverse Primer	5'-TGTAGCACGATTGCAGCATTG-3'	None
CDC	CDC-N3-P	2019-nCoV_N3 Probe	5'-FAM-AYCACATTGGCACCCGCAATCCTG-BHQ1-3'	FAM/BHQ1
CDC	RP-F	RNase P Forward Primer	5'-AGATTTGGACCTGCGAGCG-3'	None
CDC	RP-R	RNase P Reverse Primer	5'-GAGCGGCTGTCTCCACAAGT-3'	None
CDC	RP-P	RNase P Probe	5'-FAM-TTCTGACCTGAAGGCTCTGCGCG-BHQ1-3'	FAM/BHQ1
NIID	NIID-N1-F	N_Sarbeco_Forward Primer	5'-CACATTGGCACCCGCAATC-3'	None
NIID	NIID-N1-R	N_Sarbeco_Reverse Primer	5'-GAGGAACGAGAAGAGGCTTG-3'	None
NIID	NIID-N1-P	N_Sarbeco_Probe	5'-FAM-ACTTCCTCAAGGAACAACATTGCCA-TAMRA-3'	FAM/TAMRA
NIID	NIID-N2-F	NIID_2019-nCOV_N_Forward Primer	5'-AAATTTGGGGACCAGGAAC-3'	None
NIID	NIID-N2-R	NIID_2019-nCOV_N_Reverse Primer	5'-TGGCAGCTGTGTAGTCAAC-3'	None
NIID	NIID-N2-P	NIID_2019-nCOV_N_Probe	5'-FAM-ATGTCGCGCATTGGCATGGA-TAMRA-3'	FAM/TAMRA
YCH	YCH-N1-F	YCH_N1 Forward Primer	5'-CACATTGGCACCCGCAATC-3'	None
YCH	YCH-N1-R	YCH_N1 Reverse Primer	5'-GAGGAACGAGAAGAGGCTTG-3'	None

YCH	YCH-N1-P	YCH_N1 Probe	5'-FAM/ACTTCCTCA/ZEN/AGGAACAACATTGCCA-IBFQ-3'	FAM/ZEN/IBFQ
YCH	YCH-N2-F	YCH_N1 Forward Primer	5'-AAATTTGGGGACCAGGAAC-3'	None
YCH	YCH-N2-R	YCH_N1 Reverse Primer	5'-TGGCAGCTGTGTAGGTCAAC-3'	None
YCH	YCH-N2-P	YCH_N1 Probe	5'-FAM/ATGTCGCGC/ZEN/ATTGGCATGGA-IBFQ-3'	FAM/ZEN/IBFQ

NIID, National Institute of Infectious Diseases; YCH, Yamanashi Central Hospital; CDC, Centers for Disease Control and Prevention

FAM, 6-carboxyfluorescein; BHQ1, Black Hole Quencher 1; IBFQ, Iowa Black Fluorescent Quencher

Supplemental Table 2. Time course of patient symptoms, environmental cleaning, and sampling

Date	Symptoms	RT-PCR of SARS-CoV-2	Description
Before admission		Positive	Patient traveled on Diamond Princess cruise ship.
Day 1	Fever, mild cough, and pneumonia		Patient was admitted to the hospital and entered patient Room A.
Day 2	Respiratory failure		
Day 3			Patient left Room A and entered Room B.
Day 4			Patient received supplemental oxygen in Room B. Room A was cleaned.
Day 8			Five environmental samples were collected from Room A via swabbing and analyzed by RT-PCR.
Day 11		Positive in sputum	
Day 17		Negative in nasopharyngeal swab	Patient left Room B.
Day 21			Room B was cleaned. Ten environmental samples were collected from Room B via swabbing and analyzed by RT-PCR.
Day 22		Negative in nasopharyngeal swab	
Day 29		Negative in nasopharyngeal swab	
Day 35	Overall status had improved		