

Ivermectin as a potential COVID-19 treatment from the pharmacokinetic point of view

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Abstract

The broad-spectrum antiparasitic agent ivermectin has been very recently found to inhibit SARS-CoV-2 *in vitro* and proposed as a candidate for drug repurposing in COVID-19. In the present report the *in vitro* antiviral activity end-points are analyzed from the pharmacokinetic perspective. The available pharmacokinetic data from clinically relevant and excessive dosing studies indicate that the SARS-CoV-2 inhibitory concentrations are not likely to be attainable in humans.

Key words: ivermectin; SARS-CoV-2; COVID-19; Drug repurposing

Introduction

The COVID-19 pandemic has fuelled much research efforts towards repurposing of existing drugs as possible antiviral agents, whereby the therapeutic strategies have been largely based on preexisting data for the preceding coronaviral outbreaks TORS and MERS¹⁻³. The drug regulatory agencies, health authorities, key opinion leaders and policy decision makers have been significantly strained by the dilemma of evidence-based medicine and good clinical practice versus the prompt need for safe and effective treatment⁴. Unfortunately we have been witnessing huge public and political pressure for legitimation of drug-repurposing and off-label use worldwide, which nonetheless could be regarded as an acceptable compromise, pending the emergency of the current situation, but only in case of drugs with well-defined safety profiles and at least some clinical evidence in COVID-19^{4,5}. Conversely most of the treatment protocols are based on observational studies and anecdotic reports^{4,6-9}, albeit with a hope that the promptly emerging data from randomized studies will enable switching COVID-19 treatment back to the avenues of evidence-based medicine¹⁰. An exceptionally alarming phenomenon however is the public communication of drugs with preliminary *in vitro* activities against SARS-CoV-2 as

potential therapeutics for COVID-19 eventually causing malignant reverberation in social media. Such example is the otherwise very interesting study of Caly et al., recently published in *Antiviral Research*¹¹.

This paper is describing the *in vitro* antiviral activity of the antiparasitic agent ivermectin in a model of Vero/hSLAM cells infected with a SARS-CoV-2 isolate (Australia/VIC01/2020)¹¹. The authors have performed a pilot experiment using continuous exposure of the cells to ivermectin at 5 $\mu\text{mol/L}$ and found time-dependent decrease of cell associated and supernatant viral RNA. Thereafter the antiviral activity was assessed following continuous exposure to serial dilutions of ivermectin, which caused concentration-dependent antiviral effects with practically total eradication at 5 $\mu\text{mol/L}$ and half-maximal inhibition at approximately 2.5 $\mu\text{mol/L}$ ¹¹.

The academic, virological and pharmacological impact of the newly discovered antiviral effects of ivermectin against SARS-CoV-2 is beyond any doubt, but nevertheless the notion for possible clinical translation and repurposing, which has generated enormous media coverage, needs to be carefully addressed with reference to the pharmacokinetics of ivermectin. In this paper we sought to analyze the dosing regimens of the drug, the available maximal plasma concentration levels to allow detailed juxtaposition with the SARS-CoV-2 inhibitory effects and to question the paradigm for the plausibility of ivermectin repurposing in COVID-19.

Materials and methods

A literature survey was performed in order to analyze the published dose regimens and to collect human exposure data for ivermectin, following clinically relevant (150 – 800 $\mu\text{g/kg}$) or excessive dosing (up to 2000 $\mu\text{g/kg}$). The available pharmacokinetic data for ivermectin in patients with parasitic infection and healthy volunteers were pooled and the maximal plasma concentration levels (C_{max}) used as surrogates for juxtaposition with the *in vitro* SARS-CoV-2 inhibitory findings. The published concentrations shown antiviral activity were recalculated in ng/ml to allow direct comparison with the pharmacokinetic data.

Results and discussion

Ivermectin has a valuable clinical role for the management of different parasitic diseases whereby the described therapeutic regimens, could be summarized as follows: 150 $\mu\text{g/kg}$ once yearly for treatment of onchocerciasis, 200 $\mu\text{g/kg}$ as a single dose for strongyloidiasis, 150 to 200 $\mu\text{g/kg}$ twice yearly or alternatively 300 to 400 $\mu\text{g/kg}$ once yearly in endemic areas for lymphatic filariasis, and 200 $\mu\text{g/kg}$ in conjunction with topical drugs for hyperkeratotic, also known as crusted or ‘Norwegian’ scabies¹²⁻¹⁴.

Ivermectin is a semisynthetic analogue of the natural product avermectin B_{1a}, a lipophilic macrolide isolated from *Streptomyces avermitilis* developed as a crop management insecticide. Ivermectin affects

a plethora of invertebrate species, incl. parasitic nematodes, arachnids, and insects. Its mode of action on target species is by potentiating GABA-mediated neurotransmission and by binding to glutamate-gated Cl^- channels, found only in invertebrates¹³. The drug induces tonic paralysis of the musculature of susceptible parasites, and eventually death¹³. At the recommended doses, ivermectin does not readily penetrate the CNS of mammals, where GABA functions as a neurotransmitter^{13,15}. Conversely in healthy volunteers and infected patients the drug is usually well tolerated at the therapeutic dose ranges¹²⁻¹⁴. A recent meta-analysis has shown that even larger doses (up to 800 $\mu\text{g}/\text{kg}$) with a several years period of follow-up could be well tolerated in patients with parasitic infections¹⁶. The largest dose intensity with registered pharmacokinetic parameters in healthy subjects is 120 mg, corresponding to up to 2000 $\mu\text{g}/\text{kg}$ ¹².

As evident from the analyzed pharmacokinetic data both the clinically applied dosage schedules and the aforementioned excessive 120 mg dose yield blood levels at the nanogram/ml i.e. nanomolar range (Table 1). These concentrations are orders of magnitude lower, as compared to the *in vitro* antiviral end-points, described in the study of Caly et al¹¹. Table 2 summarizes the *in vitro* inhibitory concentrations, recalculated in ng/ml (based on a molecular weight of 875.1) to allow direct juxtaposition with the pharmacokinetic parameters in Table 1. Moreover the *in vitro* data have been compared to the C_{max} values, obtained after 36 mg and 120 mg doses corresponding to dose intensities of up to 700 $\mu\text{g}/\text{kg}$ ¹⁷ or 2000 $\mu\text{g}/\text{kg}$ ¹² respectively, with calculation of the corresponding exposure ratios.

The analyzed data show that at least at the clinically relevant dose ranges of ivermectin the published *in vitro* inhibitory concentrations and especially the 5 $\mu\text{mol}/\text{L}$ level causing almost total disappearance of viral RNA are virtually not achievable with the heretofore known dosing regimens in humans. The 5 $\mu\text{mol}/\text{L}$ concentration is over 50 times higher than the levels obtainable after 700 $\mu\text{g}/\text{kg}$ ¹⁷ and 17 times higher vs. the largest C_{max} found in the literature survey (247.8 ng/ml)¹². Moreover the authors' claim for achieving viral inhibition with a single dose is inappropriate because practically the infected cells have been continuously exposed at concentrations that are virtually unattainable even with excessive dosing of the drug. With other words the experimental design is based on clinically irrelevant drug levels with inhibitory concentrations whose targeting in a clinical trial seems doubtful at best.

Table 1. Published pharmacokinetic parameters for ivermectin following clinically relevant and excessive dosing

Pharmacokinetic study	Dose	Absorption parameters				Elimination	
		C_{max} (ng/ml)	C_{max} (nmol/L)	t_{max} (h)	$t_{1/2(abs)}$ (h)	$t_{1/2}$ (h)	Cl ($1 \text{ kg}^{-1} \cdot \text{day}^{-1}$)
Krishna et al., 1993 ¹⁸	6 mg (tablets)	23.1	26.4	4.3	0.5	12.6	4.28
Krishna et al., 1993 ¹⁸	12 mg (tablets)	30.4	34.7	10.3	2.5	13.4	4.03
Long et al., 2001 ¹⁹	6 mg (tablets)	20.2	23.1	4.7	1.4	11.1	7.57
Long et al., 2001 ¹⁹	12 mg (tablets)	23.5	26.9	5.3	1.4	21.1	6.53
Long et al., 2001 ¹⁹	18 mg (tablets)	31.2	35.7	5.1	1.7	16.7	10.6
Edwards et al., 1988 ²⁰	12 mg (solution)	81	92.6	3.6	–	–	–
Edwards et al., 1988 ²⁰	12 mg (tablets)	50	57.1	3.4	–	–	–
Edwards et al., 1988 ²⁰	12 mg (capsules)	46	52.6	3.6	–	–	–
Ogbuokiri et al., 1993 ²¹	150 $\mu\text{g}/\text{kg}$	37.9	43.3	–	–	–	–
Baraka et al., 1996 ²²	150 $\mu\text{g}/\text{kg}$	52.2	59.7	5.2	–	35.0	–
Elkassaby, 1991 ²³	150 $\mu\text{g}/\text{kg}$	39	44.6	5.6	–	16	–
Okonkwo, et al., 1993 ²⁴	6 mg (tablets)	38.2	43.7	4.7	–	54.5	3.1
Njoo, et al. 1995 ²⁵	150 $\mu\text{g}/\text{kg}$	-	-	-	-	19.9	–
Muñoz, et al., 2018 ¹⁷	36 mg (550 - 700 $\mu\text{g}/\text{kg}$)	96.2	109.9	3.64	-	91.77	
Guzzo et al., 2002 ¹²	90 mg	158.1	180.7	4.9	-	18.8	-
Guzzo et al., 2002 ¹²	120 mg (1404 - 2000 $\mu\text{g}/\text{kg}$)	247.8	283.2	4.2	-	19.1	-

Table 2. Comparison between the *in vitro* inhibitory concentration of ivermectin¹¹ with exemplar published C_{max} values of the drug^{12,17}

<i>In vitro</i> end-points ¹¹	Inhibitory concentrations (μmol/L)	Inhibitory concentrations recalculated in μg/ml	Inhibitory concentrations recalculated in ng/ml	Ratio between the inhibitory concentrations and clinically relevant C _{max} (96.2 ng/ml)	Ratio between the inhibitory concentrations and C _{max} after excessive dosing 247,8 (ng/ml)
IC ₅₀ - cell associated (E gene)*	2.8	2.45	2450	25.5	9.9
IC ₅₀ - supernatant (E gene)	2.4	2.1	2100	25.9	8.5
IC ₅₀ - cell associated (RdRp gene)**	2.5	2.19	2190	27.0	8.8
IC ₅₀ - supernatant (RdRp gene)	2.2	1.95	1950	24.1	7.9
Average IC ₅₀	2.475	2.166	2450	30.6	10
IC ₉₀₋₁₀₀ (highest and most effective concentration tested)	5.0	4.3755	4375	55.0	17.7

*E - envelope protein gene; ** RdRp - RNA-dependent-RNA-polymerase gene.¹¹

The repurposed antimalarial drugs hydroxychloroquine and chloroquine that have been included in numerous COVID-19 treatment protocols also have micromolar inhibitory concentrations against SARS-CoV-2²⁶ and nanomolar C_{max} values²⁷. Nevertheless these agents have enormous apparent volumes of distribution and presumably disproportionately larger tissue levels relative to the plasma concentrations, which makes the translation of the *in vitro* data plausible²⁷⁻²⁹. The other very promising agent tested in clinical trials and applied as compassionate use for COVID-19 is remdesivir^{1,30}. This broad-spectrum antiviral drug originally developed for Ebola shows potent inhibitory effects against SARS-CoV-2 with an IC₅₀ of 0.77 μmol/L (464 ng/ml)³¹. This concentration is readily attainable as the drug is given as venous infusion. The typical dosing schedule for remdesivir - initial infusion of 200 mg, followed by 100 mg/daily for a total of 5 days yields maximal plasma concentrations of 5440 ng/ml in the first day and 2610 on day 5³⁰.

In case of ivermectin, however, the potential repurposing plausibility if any is at present not very likely, because the antiviral concentrations would be attainable only after massive overdose. The therapeutic

application of ivermectin is usually not associated with significant toxicity, whereby the majority of documented adverse effects, such as: nausea, rash, dizziness, itching, eosinophilia, abdominal pain, fever, tachycardia, could be generally attributable to the gross lethality of invading microfilarias giving rise to Mazzotti-like reactions^{13,14,32}. Nevertheless at large doses the drug could penetrate the blood-brain barrier and could affect GABA-ergic transmission causing CNS depression and potentiation of the effects of benzodiazepines¹⁵. Human exposures at doses multifold higher than the therapeutic is expected to give rise to side effects similar to those documented in preclinical mammalian testing^{13,15}. Human overdoses have been associated with vomiting, tachycardia and ECG abnormalities, significant blood pressure fluctuations, CNS effects (drowsiness, ataxia) and visual disturbances (mydriasis). Accidental self-injection of a veterinary medicinal product has produced signs of clinical toxicity, albeit the drug was applied at therapeutically relevant dose (approximately 200 µg/kg)¹⁵.

It has to be emphasized that general public communication of drugs as potential COVID-19 therapeutics, based solely on *in vitro* data, is neither scientifically nor ethically appropriate. Ivermectin has been previously shown to exert antiviral activity *in vitro* against Dengue fever virus (DENV)³³, influenza virus³⁴, West Nile Virus³⁵, Venezuelan equine encephalitis virus³⁶, and heralded as possible antiviral drug, but so far there has not been any clinical translation of these data. Noteworthy a clinical trial for the treatment of Dengue fever in Thailand failed to show clinical benefits¹¹. In light to the aforementioned pharmacokinetic considerations this is not surprising given the published inhibitory concentrations against DENV1-4 ranged 1.66 - 2.32 µM³³.

Conclusion

The world has already seen epidemics of self-medication, drug shortages and even overdoses with chloroquine and hydroxychloroquine^{37,38}. Unfortunately the Caly et al. study which prompted enormous public interest has the potential to evoke similar tragic sequels, especially having in mind that in many countries the drug is only available as solutions for injection for veterinary use, whose potential for serious toxicological outcomes in humans is unanimous. In Bulgaria in particular this study has prompted the National Veterinarians` Union to share their concerns about the hysteria this study has evoked and to firmly discourage self-medication with ivermectin, which in this country is only available for use in animals³⁹. In a 10th April letter to stakeholders FDA has similarly shared its concerns on this issue and explicitly advised against any attempts for self-medication with ivermectin for COVID-19⁴⁰.

References:

1. Li G, De Clercq E. Therapeutic options for the 2019 novel coronavirus (2019-nCoV). *Nat Rev Drug Discov* 2020;19:149-50.
2. De Clercq E. Potential antivirals and antiviral strategies against SARS coronavirus infections. *Expert Rev Anti Infect Ther* 2006;4:291-302.
3. Shereen MA, Khan S, Kazmi A, Bashir N, Siddique R. COVID-19 infection: Origin, transmission, and characteristics of human coronaviruses. *Journal of Advanced Research* 2020;24:91-8.
4. Interim clinical guidance for patients suspected of/confirmed with COVID-19 in Belgium - 31 march 2020; version 6. 2020. at https://epidemiowiv-isp.be/ID/Documents/Covid19/COVID-19_InterimGuidelines_Treatment_ENG.pdf.)
5. Therapeutics and clinical trials. In: *Brigham & Women's Hospital COVID-19 Clinical Guidelines*. 2020. at <https://covidprotocols.org/protocols/04-therapeutics-and-clinical-trials#systemic-corticosteroids>.)
6. Tratamientos disponibles para el manejo de la infección respiratoria por SARS-CoV-2 (fecha de actualización: 28 de marzo de 2020). Agencia Española de Medicamentos y Productos Sanitarios (AEMPS) 2020:<https://www.aemps.gob.es/la-aemps/ultima-informacion-de-la-aemps-acerca-del-covid%E2%80%99119/tratamientos-disponibles-para-el-manejo-de-la-infeccion-respiratoria-por-sars-cov-2/?lang=en>.
7. Medicamenteuze behandelopties bij patiënten met COVID-19 (infecties met SARS-CoV-2). Stichting Werkgroep Antibiotica Beleid (SWAB) 2020:<https://swab.nl/nl/article/nieuws/494/voorlopige-behandelopties-covid-19-infecties-met-sars-cov-2>.
8. Vademecum per la cura delle persone con malattia da COVI-19. Società Italiana di Malattie Infettive e Tropicali 2020:<http://www.simit.org/medias/1569-covid19-vademecum-13-03-202.pdf>.
9. Liang T, ed. Handbook of COVID-19 Prevention and Treatment Zhejiang First Affiliated Hospital, Zhejiang University School of Medicine; 2020.
10. Thailand joins the WHO "Solidarity Trial": global testing of effective treatments of COVID-19 across 8 countries – an aggressive effort to save lives from the pandemic. World Health Organization 2020:<https://www.who.int/thailand/news/detail/20-03-2020-thailand-joins-the-who-solidarity-trial-global-testing-of-effective-treatments-of-covid-19-across-8-countries-an-aggressive-effort-to-save-lives-from-the-pandemic>.
11. Caly L, Druce JD, Catton MG, Jans DA, Wagstaff KM. The FDA-approved Drug Ivermectin inhibits the replication of SARS-CoV-2 in vitro. *Antiviral Res* 2020:104787.
12. Guzzo CA, Furtek CI, Porrás AG, et al. Safety, tolerability, and pharmacokinetics of escalating high doses of ivermectin in healthy adult subjects. *J Clin Pharmacol* 2002;42:1122-33.
13. Loukas A, Hotez PJ. Chemotherapy of helminth infections. In: Brunton LL, Lazo JS, Parker KL, eds. *Goodman & Gilman's The Pharmacological Basis of Therapeutics*, 11th Ed. New York: McGraw Hill; 2006:1073-93.
14. *British National Formulary 60*. London: BMJ Group/Pharmaceutical Press; 2010.
15. Ivermectin (INCHEM Monograph). 1992. at <http://www.inchem.org/documents/pims/pharm/ivermect.htm>.)
16. Navarro M, Camprubí D, Requena-Méndez A, et al. Safety of high-dose ivermectin: a systematic review and meta-analysis. *J Antimicrob Chemother* 2020;75:827-34.
17. Muñoz J, Ballester MR, Antonijoan RM, et al. Safety and pharmacokinetic profile of fixed-dose ivermectin with an innovative 18mg tablet in healthy adult volunteers. *PLOS Neglected Tropical Diseases* 2018;12:e0006020.
18. Krishna DR, Klotz U. Determination of ivermectin in human plasma by high-performance liquid chromatography. *Arzneimittelforschung* 1993;43:609-11.

19. Long QC, Ren B, Li SX, Zeng GX. Human pharmacokinetics of orally taken ivermectin. *Chin J Clin Pharmacol* 2001;17:203–6.
20. Edwards G, Dingsdale A, Helsby N, Orme ML, Breckenridge AM. The relative systemic availability of ivermectin after administration as capsule, tablet, and oral solution. *Eur J Clin Pharmacol* 1988;35:681-4.
21. Ogbuokiri JE, Ozumba BC, Okonkwo PO. Ivermectin levels in human breastmilk. *Eur J Clin Pharmacol* 1993;45:389-90.
22. Baraka OZ, Mahmoud BM, Marschke CK, Geary TG, Homeida MM, Williams JF. Ivermectin distribution in the plasma and tissues of patients infected with *Onchocerca volvulus*. *Eur J Clin Pharmacol* 1996;50:407-10.
23. Elkassaby MH. Ivermectin uptake and distribution in the plasma and tissue of Sudanese and Mexican patients infected with *Onchocerca volvulus*. *Trop Med Parasitol* 1991;42:79-81.
24. Okonkwo PO, Ogbuokiri JE, Ofoegbu E, Klotz U. Protein binding and ivermectin estimations in patients with onchocerciasis. *Clin Pharmacol Ther* 1993;53:426-30.
25. Njoo FL, Beek WM, Keukens HJ, et al. Ivermectin detection in serum of onchocerciasis patients: relationship to adverse reactions. *Am J Trop Med Hyg* 1995;52:94-7.
26. Colson P, Rolain JM, Lagier JC, Brouqui P, Raoult D. Chloroquine and hydroxychloroquine as available weapons to fight COVID-19. *Int J Antimicrob Agents* 2020:105932.
27. Thummel KE, Shen DD, Isoherannen N, Smith HE. Appendix II. Design and optimization of dosage regimens. In: Brunton LL, Lazo JS, Parker KL, eds. *Goodman & Gilman's The Pharmacological Basis of Therapeutics*, 11th Ed. New York: McGraw Hill; 2006:1787-888.
28. Shapiro TA, Goldberg DE. Chemotherapy of protozoal infections: malaria. In: Brunton LL, Lazo JS, Parker KL, eds. *Goodman & Gilman's The Pharmacological Basis of Therapeutics*, 11th Ed. New York: McGraw Hill; 2006:1021-47.
29. Yao X, Ye F, Zhang M, et al. In Vitro Antiviral Activity and Projection of Optimized Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). *Clinical Infectious Diseases* 2020.
30. European Medicines Agency /178637/2020/Human Medicines Division. Summary on compassionate use - Remdesivir Gilead, International Nonproprietary Name: remdesivir, Procedure No. EMEA/H/K/5622/CU (03 Apr. 2020). 2020. at https://www.ema.europa.eu/en/documents/other/summary-compassionate-use-remdesivir-gilead_en.pdf.)
31. Wang M, Cao R, Zhang L, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res* 2020;30:269-71.
32. Fawcett RS. Ivermectin use in scabies. *Am Fam Physician* 2003;68:1089-92.
33. Tay MYF, Fraser JE, Chan WKK, et al. Nuclear localization of dengue virus (DENV) 1–4 non-structural protein 5; protection against all 4 DENV serotypes by the inhibitor Ivermectin. *Antiviral Research* 2013;99:301-6.
34. Götz V, Magar L, Dornfeld D, et al. Influenza A viruses escape from MxA restriction at the expense of efficient nuclear vRNP import. *Scientific Reports* 2016;6:23138.
35. Yang SNY, Atkinson SC, Wang C, et al. The broad spectrum antiviral ivermectin targets the host nuclear transport importin alpha/beta1 heterodimer. *Antiviral Res* 2020:104760.
36. Lundberg L, Pinkham C, Baer A, et al. Nuclear import and export inhibitors alter capsid protein distribution in mammalian cells and reduce Venezuelan Equine Encephalitis Virus replication. *Antiviral Res* 2013;100:662-72.
37. Nigeria records chloroquine poisoning after Trump endorses it for coronavirus treatment. 2020. at <https://edition.cnn.com/2020/03/23/africa/chloroquine-trump-nigeria-intl/index.html>.)

38. Jakhar D, Kaur I. Potential of chloroquine and hydroxychloroquine to treat COVID-19 causes fears of shortages among people with systemic lupus erythematosus. *Nature Medicine* 2020.
39. Veterinarians warn: do not buy ivermectin for treatment of humans, it is very toxic. 2020. at <https://news.bnt.bg/news/veterinari-preduprezhdavat-ne-kupuvaite-ivermektin-za-lechenie-na-hora-silno-toksichen-e-1047797news.html>.)
40. FDA letter to stakeholders: Do not use ivermectin intended for animals as treatment for COVID-19 in humans (April 10, 2020). 2020. at <https://www.fda.gov/animal-veterinary/product-safety-information/fda-letter-stakeholders-do-not-use-ivermectin-intended-animals-treatment-covid-19-humans>.)