Estimation of Rift Valley fever virus spillover to humans during the Mayotte 2018-2019 epidemic

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28 Abstract

29 Rift Valley fever (RVF) is an emerging, zoonotic, arboviral haemorrhagic fever threatening livestock and humans mainly in Africa. RVF is of global concern, having expanded its 30 geographical range over the last decades. The impact of control measures on epidemic dynamics 31 32 using empirical data has not been assessed. Here, we combined seroprevalence livestock and 33 human RVF case data from the 2018-2019 epidemic in Mayotte, with a dynamic mathematical model. Using a Bayesian inference framework, we estimated viral transmission potential amongst 34 35 livestock, and spillover from livestock to humans, through both direct contact and vector-mediated 36 routes. Model simulations were used to assess the impact of vaccination on reducing the human 37 epidemic size. Reactive vaccination immunising 20% of the livestock population reduced the 38 number of human cases by 30%. To achieve a similar impact, delaying the vaccination by one 39 month required using 50% more vaccine doses, and vaccinating only humans required 20 times 40 as more as the number of doses for livestock. Finally, with 53.92% (95%Crl [44.76-61.29]) of 41 livestock estimated to be immune at the end of the epidemic wave, viral re-emergence in the next 42 rainy season (2019-2020) was unlikely. We present the first mathematical model for RVF fitted to 43 real-world data to estimate virus transmission parameters, and able to inform potential control programmes. Human and animal health surveillance, and timely livestock vaccination appear to 44 45 be key in reducing disease risk in humans. We furthermore demonstrate the value of a One 46 Health quantitative approach to surveillance and control of zoonotic infectious diseases.

47 Introduction

Controlling zoonotic and vector-borne infections is complex, as it requires an accurate 48 49 understanding of pathogen transmission within animal populations, and pathogen spillover to 50 humans, whilst accounting for environmental factors affecting vector population dynamics (1.2). 51 Rift Valley fever (RVF) is an emerging zoonotic arbovirosis causing haemorrhagic fever. RVF is a 52 threat for both animal and human health, mainly in Africa (3). Livestock (cattle, sheep and goats) 53 are RVF virus amplifying hosts, acquiring infection through the bites of infectious mosquitoes 54 (mainly Aedes spp. and Culex spp.) (4). Humans get infected by direct contact with infectious 55 animal tissues (upon abortions or animal slaughter), although vector transmission may also play a 56 role (4,5). Since 2015, RVF has been listed as a priority emerging disease by the WHO R&D 57 Blueprint (6). A major concern is the expansion of its geographical range over recent decades (5,7). Current disease control options for reducing disease risk in humans heavily rely on 58 59 controlling virus transmission in animal populations. The impact of disease control measures in livestock on reducing RVF risk in humans has not yet been assessed, and doing so requires 60 61 estimating key transmission parameters between livestock, and from livestock to humans; using 62 animal and human epidemiological data.

63 Mayotte, an island located in the South Western Indian Ocean region, reported a RVF epidemic in 2007-2008 (8). In a previous paper, we used longitudinal livestock seroprevalence data to 64 model RVF virus emergence in the livestock population, and we estimated that the likelihood of 65 re-emergence was very low in a closed ecosystem (i.e. without introduction of infectious animals). 66 However, a few imported infectious animals could trigger another large epidemic, as the herd 67 immunity declined due to livestock population turnover (9). In 2018, about ten years after the 68 69 previous epidemic, RVF outbreaks were reported in several East African countries (e.g. Kenya, 70 South Sudan, Uganda, Rwanda) (10,11). In Mayotte, between November 2018 and August 2019, 71 a total of 143 human cases (RVF virus RT-PCR confirmed) were reported (Fig. 1A). The virus 72 belongs to the Kenya-2 clade (12), which is closely related to the strains detected in recent 73 outbreaks in Eastern Africa. The Veterinary Services of Mayotte, the regional health authorities 74 (Agence de Santé Océan Indien) and the French Public Health Agency (Santé Publique France) 75 did further epidemiological investigations to assess temporal patterns in occurrence of the 76 infection in the animal population, and to identify possible routes of human exposure to RVF 77 virus. These investigations generated a uniquely well documented RVF epidemic dataset, 78 including RVF seroprevalence and incidence data in animal and humans.

79 We present these data and use them to extend and fit a mathematical model of RVF virus 80 transmission in livestock (9), and explicitly account for viral spillover from livestock into the human

81 population. We fit this model simultaneously to the infection patterns in livestock and human

observed during the 2018-2019 epidemic, allowing for the first time, (i) to estimate the level of

83 RVF virus transmission amongst livestock and spillover from livestock to humans by both direct

so contact and vector-mediated routes, (ii) to estimate the likelihood of another epidemic the

following year, and (iii) to assess the impact of potential vaccination strategies in livestock and

86 humans on reducing disease occurrence in humans.

87 <u>Results</u>

88 The course of the epidemic in livestock and humans

Between November 2018 and August 2019, 143 RVF human cases were reported. The epidemic peaked mid-February (February 11-17, 2019), with 18 weekly confirmed cases, six to seven weeks following the rainfall peak (Fig. 1A). About two-third of investigated cases reported a direct contact with livestock or its tissues (incl. milk consumption) (68%, n=86), whilst 32% (n= 41) reported no previous contact with animals (Fig 1A. cases in red and green, respectively).

94 Livestock sera (n=1,169) collected by the Veterinary Services between July 2018 and June 2019 95 were tested against RVF IgG. To assess the timing of emergence of the virus in the livestock 96 population, we plotted guarterly age-stratified RVF IgG prevalence, using only tested animals for 97 which the date of birth was available (n=493). In July - September 2018, that is before the report 98 of the first human case, most seropositive animals were in the oldest age groups (Fig. 1D), 99 possibly indicating viral exposure during the previous re-emergence (9). The IgG seroprevalence 100 increased in all age groups in January-March (Fig. 1E), and then in April-June 2019 (Fig. 1F), 101 evidencing that the emergence of the virus in the livestock population, was coincident with the 102 report of cases in humans.

103 Ongoing viral phylogenetic analyses on human derived-samples (12), and IgM positive livestock 104 seized from informal trade between June and August 2018 (Table S1), suggest that the virus was 105 likely introduced from Eastern Africa into Mayotte between June and August 2018, through the 106 movements of infectious animals.

107 Epidemic model

108 We modelled virus transmission amongst livestock as a function of rainfall, therefore varying 109 along the study period. RVF virus spillover from livestock to humans was modelled by both direct 110 contact, assuming a time-invariant transmission rate, and vector-mediated transmission, defined 111 as a function of rainfall (see Methods).

112 Transmission parameters. By fitting this model to the RVF datasets, the time-varying 113 reproduction number in the livestock population was estimated to peak at $R_s(t)=1.87$ (95%) 114 Credible Interval Crl [1.53-2.69]), in the second half of January (January 14-27, 2019) (Fig. S3), two weeks following the rainfall peak, and three to four weeks prior to the predicted epidemic 115 116 peaks in livestock and humans (Fig. 2A). This corresponded to a transmission rate amongst livestock ($\beta_{L-L}(t)$) at 8.92 per 100,000 livestock heads per day (95%Crl [1.09-7.77]). The 117 spillover rate from livestock to humans by direct contact (β_{L-H}^{C}) was estimated to 1.78 per 10 million persons per day (95%Crl [1.29 – 2.61]) (Table S2), and the maximum values of the time-varying spillover transmission rate ($\beta_{L-H}^{V}(t)$) was 1.33 per 10 million persons per day (95%Crl 118 119 120 121 [2.25-8.46]).

Model predictions. Using the estimated parameters, the simulated number of human reported cases was 181 (95% CrI [138-233]), with two third resulting from direct contact (n=111, 95%CrI [80-149]) and one third from vector transmission (n=70, 95%CrI [44-102]) (Table 1), in agreement with the observed data (Fig. 1B-1C). The predicted age-stratified IgG seroprevalence in livestock between January and June 2019 were in good agreement with the observed data as well (Fig. 1E-1F). The simulated incidence in livestock cases peaked mid-February (February, 11-17),

concomitantly with the peak in human vector-mediated transmission, whilst the number of human
cases by direct contact reached its maximum values one week later (February, 18-24) (Fig. 2A).
Finally, by the end of the epidemic wave, 18,460 (95%Crl [14,926-21,154]) animals were affected
resulting in 53.92 % (95%Crl [44.76-61.29]) of the livestock population being immune (Fig. 2B
and Table 1). The overall predicted number of human cases (both reported and not reported) was
estimated to have reached 9,566 (95%Crl [7,793-11,772]), resulting in 3.73% (95%Crl [3.034.56]) of the human population being immune (Table 1).

135 In this setting, the likelihood of virus re-emergence the following rainy season (2019-2020) was 136 less than 2.5% (Fig. 2A), with the time-varying effective reproductive number $R_e(t)$ falling below 137 unity following the epidemic peaks and remaining very close to or below unity over the second 138 year of the simulations (Fig. 2C).

139 Vaccination scenarios. Probabilistic forecasts were also used to assess the impact of different 140 livestock and humans vaccination strategies on the size of the epidemic in both animals and 141 humans (Fig 3A-3D and Table 1). A reactive and mass vaccination campaign in livestock immediately after the report of the first human case (i.e. 6,000 doses in December 2018) allowed 142 143 a reduction in the epidemic size by a third (median number of humans cases = 113 cases, 144 median number of livestock cases = 11,447), while waiting one more month would have required 145 50 % more vaccine doses to achieve a similar impact (9,000 doses in January 2019, median 146 number of humans cases = 115, median number of livestock cases = 11,573). Finally, a 147 vaccination programme targeting only humans would require immunising half of the population 148 (128,250 doses in December 2018) to reduce similarly the number of human cases (median=115 149 cases), whilst, of course, not impacting on the number of livestock cases.

150 Discussion

We present the first RVF epidemic dataset combining both livestock and human surveillance data, and use it to parameterise a mathematical model. We estimated, for the first time, transmission rates amongst livestock and spillover to humans using empirical epidemic data. This also allowed the quantitative assessment of the importance of timely livestock vaccination in reducing disease risk in humans during an epidemic, useful to inform potential control programmes, and illustrating the importance of One Health surveillance in the management of zoonotic diseases.

158 The IgM testing of illegally imported livestock suggested that the virus may have been introduced 159 in Mayotte around June-August 2018, which is in agreement with the timing of RVF outbreaks on the East African mainland (11) and corresponds to the dry season in Mayotte. Viral transmission 160 161 might have been maintained on the island at a low level in the dry season, or the virus might have 162 been several times introduced, and the epidemic started only following the start of the rainy 163 season (that is in October). The epidemic is likely to have therefore resulted from a recent viral 164 re-introduction, rather that viral persistence over the last ten years, as concluded about the 2007-165 2008 epidemics, in a previous study (9).

166 The systematic testing by RT-PCR of humans showing dengue-like syndrome performed in 167 Mayotte for the last ten years (since 2008), provides additional evidence that RVF had been 168 absent from the Island for a decade, and that the presented epidemic curve accurately reflected its actual timing. During the epidemic, mitigation strategies such as vector control around houses 169 170 of human cases (i.e. post-detection) and the diffusion of prevention messages on milk 171 consumption and exposure to animals were communicated, from February 27th onwards (13), that 172 is two weeks after the peak. Therefore, these measures are likely to have had a moderate impact 173 on the epidemic size, whilst not affecting the time of the epidemic peak. In addition, the timing of 174 the epidemic was corroborated by the observed changes in livestock seroprevalence, exhibiting a 175 clear pattern of viral emergence. Most livestock sera (90%) were collected and tested as part of 176 the regular annual surveillance campaign. As 10% of these samples were collected in areas reporting human cases, the proportion of seropositive animals may have been overestimated. 177 178 However, most animal sampling was conducted from January 2019 onwards, when RVF virus

had already spread across the whole island. In addition, our model predicted that 53.92 %
(95%Crl [44.76-61.29]), of the livestock population was immune at the end of the simulated
epidemic wave (August 2019), which was in line with estimates from the previous emergence in
2007-2008 (9).

Previous RVF models parameterised the transmission rate from livestock to humans by direct 183 184 contact as an input parameter at 1.7 per 10 thousand persons per day (14-16). The 185 epidemiological investigations conducted in this epidemic assessed whether human cases 186 reported a direct contact with animals or their infectious tissues, and human cases without prior 187 contact with such materials. This allowed for the first time estimating both RVF virus spillover to 188 humans by direct contact and by vector transmission from epidemic data. These estimated 189 transmission rates can be used as a benchmark for further modelling work. RVF human cases 190 with or without previous contact with animals or animal products have been reported in other 191 settings (17,18). Here, the reported fraction of cases without previous animal contact (32%) was 192 three times higher than in South Africa (10%) (17). Several reasons may explain these 193 differences. For example, this could result from a recall bias from people interviewed in Mayotte, 194 or from the fact that in South Africa, people were tested following reports of RVF in animals (17).

195 Rainfall is a known driver for RVF virus transmission (19) and was used as a proxy for vector 196 abundance. We assumed a 14-days lag between rainfall and its impact on vector abundance 197 based on previous modelling studies on RVF vectors population dynamics (20,21). Temperature 198 above 26°C may also promote RVF virus transmission (22-24). The temperatures of Mayotte 199 varying annually between 25°C and 35°C (9), we assumed that in this specific setting, 200 temperature would not be a major driver for viral transmission. In areas with cooler temperatures, 201 such as South Africa (25), temperature may need to be taken into account (26). The highest 202 estimated $R_s(t)$ value was 1.87 (95%Crl [1.53-2.69]), yet in line with previous estimations of R_o 203 (14,27,28). The baseline model, with constant transmission parameters, had a similar DIC than the rainfall-dependent model, and showed R_0 values within the same range. Whilst this may 204 205 suggest a smaller influence of environmental factors on the RVF viral transmission dynamics 206 during this epidemic, it may also suggest that upon the conditions met for emergence -(i) the 207 presence of the virus. (ii) a susceptible livestock population and (iii) the presence of vectors - the 208 epidemic fade-out likely resulted from a depletion of susceptible livestock. This was corroborated 209 by the small likelihood of re-emergence in the following rainy season, with an effective 210 reproductive number $R_{e}(t)$ remaining close or below unity in the months following the end of the 211 2018-2019 epidemic, due to the high proportion of immune animals.

212 A limitation of the model was that the reporting rate in humans was unknown, and defined based on data from the 2007-2008 epidemic (29). This relied on the assumption that both the 2007-2008 213 214 and 2018-2019 epidemics affected the same number of people. Whilst there is no data available 215 on human infection patterns to support this assumption, our previous work estimated a post-216 epidemic livestock seroprevalence (9) which was similar to our current estimates, supporting the 217 assumption that both epidemic sizes may be comparable. Further data collection estimating 218 human post-epidemic seroprevalence would allow an accurate estimation of this reporting rate. 219 Finally, the livestock model was built with similar assumptions than in our previous paper (9). This 220 included a latent (E) and an infectious (I) period of 7 days in livestock, accounting for the extrinsic 221 incubation period in the vector (3-7 days), and the latent (1-6 days) and infectious stages (3-6 222 days) in livestock (30-33), without explicitly modelling these processes. Although this may have 223 slightly impacted on the predicted timing of the epidemic peak in humans, our model predictions 224 were in agreement with the observations. In addition, this did not impact on the fitting with 225 livestock data, as we fitted on the (R) compartment, aggregating data over three-month periods. 226 We also assumed homogeneous mixing. Mayotte is a small island (374km²), the ecosystem 227 shows limited spatial variations, livestock production systems are extensive with animals raised 228 outdoor year round (9), compatible with the assumption that the livestock population was equally 229 exposed to RVF mosquito vectors. Accounting for spatial heterogeneity, and testing for finer 230 vaccination protocols would have required the use of epidemic data at a smaller spatio-temporal

resolution. Our model can however be expanded into a metapopulation structure, and parameters
 further refined, in ecosystems with epidemic data available at finer spatial and temporal scales.

233 The impact and cost-effectiveness of livestock vaccination has been assessed in specific RVF 234 high-risk areas in Kenya using simulation modelling (32,33). Instead, our analysis allows 235 predicting the impact of vaccination strategies on reducing the number of human and animal 236 cases, through a model calibrated from epidemic data. Our findings provide evidence that 237 reactive animal vaccination is the most effective control measure, preventing both human and 238 livestock cases, and requiring a smaller number of vaccine doses. The characteristics of the 239 vaccine used in the vaccination scenarios (highly immunogenic, single dose, and safe) were 240 those targeted by WHO R&D Blueprint (34), and not the existing ones. In practice, currently 241 available RVF vaccines have different immunogenic and safety characteristics, with some of them 242 requiring boosters (35), and the choice of which vaccine to use on the field may vary upon the 243 epidemiological context. In addition, during this epidemic, livestock were not vaccinated due the 244 absence of a vaccine with a EU marketing authorisation (Mayotte is an EU outermost region) 245 (36). However, we highlight the importance of the development of contingency planning, 246 availability of emergency funds and a suitable vaccine.

247 In conclusion, we have presented a uniquely detailed investigation into an outbreak of an 248 emerging arbovirus, combining animal and human data, with a mathematical model for RVF. 249 Early detection and rapid vaccination are critical to RVF control at the early stage of the epidemic. 250 Disease surveillance in animals, contingency planning, and the timely implementation of livestock 251 vaccination, are key for reducing human disease risk. This work represents a collaboration 252 between public health agency, animal health surveillance network, farmers' association, and 253 researchers, initiated from the start of the epidemic, and conducted as a collaborative work as the 254 epidemic unfolded. Delays in getting livestock data were inherent to climatic conditions (storms) 255 and field work constraints in remote areas. Nevertheless, we addressed in practice the 256 challenges of a quantitative One Health approach (37), and illustrated its value to surveillance and 257 control of zoonotic emerging infectious diseases. Our model can be further expanded, refined and 258 recalibrated for other ecosystems.

259 Materials and Methods

260 **RVF datasets**

Human data. Human incident case data were collected from patients showing dengue-like
symptoms and consulting a GP, and who subsequently tested positive for RVF virus RT-PCR
(38). Cases were interviewed using a structured questionnaire administered by Sante Publique
France health epidemiologists (39). The number of incident cases was aggregated by week.

265 Livestock data. During the study period, livestock sera were sampled by field veterinarians 266 according to two protocols: RVF targeted surveys around human cases and the regular annual 267 surveillance campaign (SESAM) which is implemented since 2008 (8). The sera from the RVF 268 targeted surveys were collected around human cases and were collected between January and 269 March only. However, due to the rapidly increasing number of human cases and logistics 270 constraints, the Veterinary Services instead requested field veterinarians to sample animals from 271 the annual surveillance campaign only, depending on their regular field visits, therefore not 272 depending on human cases. In total, between July 2018 and June 2019, a total of 1,169 livestock 273 sera were collected (842 from the annual surveillance and 146 from human investigations), and 274 tested against RVF IgG (ID Screen RVF Competition ELISA, IDVet, Grabels, France, Se=97 %, 275 Sp=100 % (40)). Date of birth was available for 493 of these sampled animals (with 9% from RVF 276 targeted surveys, Table S3). In order to follow the emergence of the virus in the livestock 277 population over a year, we plotted quarterly age-stratified RVF IgG prevalence (Fig. 1D-1F).

Origin of the virus. To investigate the possible time window of virus introduction from imported infected animals, we collated serological data from illegally imported livestock seized by the

Veterinary Services between June and August 2018. These animals were tested RVF IgM by
 ELISA (indicative of recent infections) (Table S1).

282 Epidemic model

We modelled RVF epidemic from the start of the rainy season, the first week of October 2018 (October, 1-7), one month prior to the report of the first human case, up to the first week of August 2019 (July, 29-August, 4). No more human cases were reported after this date.

286 Transmission amongst livestock. We adapted the previously developed SEIR model of RVF 287 virus transmission amongst Mayotte livestock (9) to the current epidemiological context. For full 288 details on the model structure and equations, see Metras et al. 2017 (9). We kept the previous underlying demographic livestock population age-structure (10 yearly age-groups) for fitting 289 purpose, and we used a discrete-time deterministic framework, with a daily time step. In the 290 previous model, the transmission parameter amongst livestock ($\beta_{L-L}(t)$) and corresponding time-291 varying reproductive number $(R_s(t))$ were assumed to be vector-borne and modelled as a function 292 293 of monthly NDVI (Normalized Difference Vegetation Index) values, as a proxy for vector 294 abundance. Here, instead of using monthly NDVI, we used rainfall data (41) at a daily time step, 295 since the model time step and the human epidemic curve available for fitting had a smaller time 296 resolution. We also included a lag of 14 days between rainfall and its impact on the vector 297 abundance (21,22). To look at the temporal pattern of the viral transmission over time, we also 298 calculated $R_e(t)$, the time-varying effective reproduction number, as the product of $R_s(t)$ with the 299 proportion of susceptible livestock at time t (SI Methods).

300 Spillover into humans. We added a module simulating RVF virus transmission from livestock 301 into the human population. We assumed that susceptible humans (S_H) became infected following exposure with infectious livestock by direct contact (E_{H}^{c}) at a constant rate β_{L-H}^{c} , and by vector-302 mediated route (E_{H}^{V}) at a time-varying rate $m{eta}_{L-H}^{V}(t)$, scaled on the rainfall-dependent within-303 livestock transmission ($\beta_{L-L}(t)$). Infected individuals E_H^C and E_H^V successively moved to their 304 respective infectious states (I_{H}^{C} and I_{H}^{V}); after which they moved into the immune compartment (305 306 $R_{\rm H}$) (Fig. S1), assuming they remained immune until the end of the study period. The model 307 equations, transmission parameters and the formulation of the forces of infection from livestock 308 $\lambda_{contact}(t)$ and vectors $\lambda_{vector}(t)$ are presented in **SI Methods**.

309 Parameterisation and model fitting. Input parameters were those related to the natural history 310 of infection and demographics in both livestock and human populations (Table S4). The proportion of immune animals at t_0 was informed from the aggregated July-September 2018 IgG 311 livestock seroprevalence campaign (Fig. 1D). The reporting fraction of human cases was set to 312 313 ρ =1.9%, as a post-epidemic serological study in humans, conducted in 2011 in Mayotte, 314 estimated that 3.5% (95%CI [2.6-4.8]) of the human population was RVF IgG-positive (30). Assuming a population size of 212,645 inhabitants in 2012 (42), this corresponded to an average 315 316 of 7,442 persons being seropositive. Assuming that the sizes of the 2007-2008 and 2018-2019 317 epidemics were similar, the detection of 143 cases in the 2018-2019 epidemics suggests a 318 reporting fraction of 1.9% (95% CI [1.4-2.6]). Finally, input rainfall data were downloaded from the 319 Meteofrance website, as cumulated rainfall over 10-day periods (41). Daily rainfall was calculated 320 by dividing these values by ten over each 10-day period.

321 Five parameters were estimated by fitting the model to the human and livestock epidemic data (Table S3). Two parameters related to the rainfall-dependent transmission amongst livestock (A 322 and *B*), two parameters estimated the spillover to humans, via contact with livestock (β_{L-H}^{C}) and 323 via vector (scaling factor X), and the fifth parameter was the number of infectious livestock at t_0 324 325 (I_{ivo}) . Parameter estimation was done by fitting simultaneously the (i) quarterly age-stratified 326 simulated proportion of immune livestock ($p_{a,q}$) to quarterly RVF IgG prevalence (Fig. 1E-1F); (ii) 327 the simulated weekly number of reported incident cases in humans by direct contact (Fig. 1B) and 328 (iii) the simulated weekly number of reported incident cases in humans by vector-mediated route

(Fig. 1C), to the observed weekly number of reported cases via both transmission routes. Values of those five parameters were sampled from their prior distribution $\theta = \left[A, B, \beta_{L-H}^{C}, X, I_{liv0}\right]$ using a Monte Carlo Markov Chain Metropolis-Hastings (MCMC-MH) algorithm, implemented in the fitR package (43). Finally, to assess the impact of rainfall over the course of the epidemic, we also fitted a baseline model for which all transmission parameters were constant over time (Table S5). Details on models equations, parameter estimation, model fitting, and model comparison are presented in *SI Methods*.

336 Forecasting and vaccination scenarios. We did probabilistic projections for seven scenarios 337 (Table 1). For all scenarios, we simulated 2,500 stochastic trajectories by sampling randomly parameter values from the joint posterior distribution. Scenario 1 aimed at estimating the 338 339 likelihood of virus re-emergence, without disease control intervention, in the following rainy 340 season (in 2019-2020), in a closed ecosystem, using the same rainfall data as during the 2018-341 2019 rainy season. Scenarios 2-6 aimed at assessing the impact that different livestock 342 vaccination strategies could have had on the number of human and livestock cases during the 343 2018-2019 epidemic. We assumed the use of a single-dose highly immunogenic vaccine (90 % 344 vaccine efficacy) (34,35), and a 14-days lag between vaccination and build-up of immunity. 345 Figures of vaccination campaigns in Mayotte in 2017 (against blackleg, a livestock disease), 346 showed that about 3,000 vaccine doses are routinely administered to livestock over a year by 347 local veterinarians. Scenario 2 tested the impact of administrating all these 3,000 doses in one month, in December 2018, immediately after the report of the first human case (joint animal-348 349 human alert date for response), corresponding to the current vaccinating capacity in Mayotte in 350 an emergency setting. Scenario 3 assumed an extra-vaccine supply and an emergency mass 351 vaccination, allowing 6,000 doses to be administered in December 2018. We also assessed the 352 impact of vaccinating livestock in January 2019, one month following the report of first human 353 case, allowing extra time for organising the vaccination campaign: 3,000 doses (Scenario 4), 354 6,000 doses (Scenario 5) and 9,000 doses (Scenario 6). Finally, to assess the impact of a 355 reactive and mass vaccination only in humans, we simulated a 50% vaccination coverage of the 356 human population in December 2018 (i.e. 128,250 doses) (Scenario 7). Vaccination equations 357 and diagram are presented in SI Methods and Fig. S2.

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367 Author Contributions

RM, WJE, LD, YH, MS conceptualized and designed the study. CY, LD, SC, YH, MS collated the
data and did data management. RM, GF performed the analyses. RM, WJE, CY, LD, GF, AC, SF,
GLG, CS, EC, LF, YH, MS interpreted and discussed the data and results. RM was responsible
for drafting the manuscript. All authors reviewed and approved the final manuscript.

372 **References**

1. M E. J. Woolhouse, C. Dye, Preface. *Philos. Trans. Roy. Soc. London Ser B.* 356, 981–982
(2001).

2. B. A. Jones, D. Grace, R. Kock, S. Alonso, J. Rushton, M. Y. Said, D. McKeever, F. Mutua, J.
Young, J. McDermott, D. U. Pfeiffer, Zoonosis emergence linked to agricultural intensification and
environmental change. *Proc. Natl. Acad. Sci. U. S. A.* **110**, 8399-8404 (2013).

378 3. M. H. A. Clark, G. M. Warimwe, A. Di Nardo, N. A. Lyons, S. Gubbins, Systematic literature 379 review of Rift Valley fever virus seroprevalence in livestock, wildlife and humans in Africa from 380 1968 to 2016. *PLoS Negl. Trop. Dis.* **12**, e0006627 (2018).

4. B. H. Bird, T. G. Ksiazek, S. T. Nichol, N. J. MacLachlan, Rift Valley fever virus. *J. Am. Vet. Med. Assoc.* 234, 883–893 (2009).

5. M.O. Nanyingi, P. Munyua, S. G. Kiama, G. M. Muchemi, S. M. Thumbi, A. O. Bitek, B. Bett, R.
M. Muriithi, M. K. Njenga, A systematic review of Rift Valley Fever epidemiology 1931-2014. *Infect. Ecol. Epidemiol.* 5 (2015).

386 6. World Health Organization. Epidemic and pandemic-prone diseases, List of Blueprint priority
 387 diseases. Available at <u>http://www.emro.who.int/fr/pandemic-epidemic-diseases/news/list-of-</u>
 388 <u>blueprint-priority-diseases.html</u> Accessed on 8 March 2020 (2018).

7. R. Hatchett, N. Lurie, Outbreak responses as an essential component of vaccine development.
 Lancet. Infect. Dis. 19, e399-e403 (2019).

8. R. Métras, L. Cavalerie, L. Dommergues, P. Mérot P, W. J. Edmunds, M. J. Keeling, C. CêtreSossah, E. Cardinale, The Epidemiology of Rift Valley Fever in Mayotte: Insights and
Perspectives from 11 Years of Data. *PLoS Negl. Trop. Dis.* **10**, e0004783 (2016).

9. R. Métras, G. Fournié, L. Dommergues, A. Camacho, L. Cavalerie, P. Mérot, M. J. Keeling, C.
Cêtre-Sossah, E. Cardinale, W. J. Edmunds, Drivers for Rift Valley fever emergence in Mayotte:
A Bayesian modelling approach. *PLoS Negl. Trop. Dis.* **11**, e0005767 (2017).

397 10. ProMED. Rift Valley fever - Kenya (02): (Wajir). Published Date: 2018-06-09. Archive
 398 Number: 20180609.5847216. Accessed on 05 December 2019 (2018).

11. Food and Agriculture Organization of the United Nations. EMPRES-i. Global Animal Disease
 Information System. Available at : <u>http://empres-i.fao.org/eipws3g/</u> Accessed on 11 December
 2019.

402 12. A. Kwasiborski, L. Collet, V. Hourdel, M. Vandenbogaert, C. Batejat, J. C. Manuguerra, J.
403 Vanhomwegen, V. Caro. Molecular investigation of Rift Valley Fever outbreak in Mayotte, 2018.
404 Available at: <u>https://programme.europa-organisation.com/slides/programme_ricai-2019/CO-</u>
405 <u>095.pdf</u>, Accessed on 20 March 2020 (2019)

406 13. ProMED. Rift Valley fever - Mayotte (12): human, cattle. Published Date: 2019-05-28. Archive
407 Number: 20190528.6489852. Accessed on 09 March 2020 (2019).

408 14. L. Xue, M. H. Scott, L. W. Cohnstaedt, C. Scoglio, A network-based meta-population 409 approach to model Rift Valley fever epidemics. *J Theor. Biol.* **306**, 129–144 (2012).

410 15. S. C. Mpeshe, H. Haario, J. M. Tchuenche, A Mathematical Model of Rift Valley Fever with 411 Human Host. *Acta Biotheor.* **59**, 231 (2011).

9

412 16. J. Lugoye, J. Wairimu, C. B. Alphonce, M. Ronoh, Modeling Rift Valley fever with treatment 413 and trapping control strategies. *Appl. Math.* **7**, 556-568 (2016).

414 17. B. N. Archer, J. Thomas, J. Weyer, A. Cengimbo, D. E. Landoh, C. Jacobs, S. Ntuli, M.
415 Modise, M. Mathonsi, M. S. Mashishi, P. A. Leman, C. le Roux, P. J. van Vuren, A. Kemp, J. T.
416 Paweska, L. Blumberg, Epidemiologic Investigations into Outbreaks of Rift Valley Fever in
417 Humans, South Africa, 2008-2011. *Emerg. Infect. Dis.* **19**, 1918–1925 (2013).

18. T. R. Shoemaker, L. Nyakarahuka, S. Balinandi, J. Ojwang, A. Tumusiime, S. Mulei, J.
Kyondo, B. Lubwama, M. Sekamatte, A. Namutebi, P. Tusiime, F. Monje, M. Mayanja, S.
Ssendagire, M. Dahlke, S. Kyazze, M. Wetaka, I. Makumbi, J. Borchert, S. Zufan, K. Patel, S.
Whitmer, S. Brown, W. G. Davis, J. D. Klena, S. T. Nichol, P. E. Rollin, J. Lutwama, First
Laboratory-Confirmed Outbreak of Human and Animal Rift Valley Fever Virus in Uganda in 48
Years. *Am. J. Trop. Med. Hyg.* 100,659–671 (2019).

424 19. R. Sang, J. Lutomiah, M. Said, A. Makio, H. Koka, E. Koskei, A. Nyunja, S. Owaka, D. 425 Matoke-Muhia, S Bukachi, J. Lindahl, D. Grace, B. Bett, Effects of Irrigation and Rainfall on the 426 Population Dynamics of Rift Valley Fever and Other Arbovirus Mosquito Vectors in the Epidemic-427 Prone Tana River County, Kenya. *J. Med. Entomol.* **54**, 460–470 (2017).

20. C. Talla, D. Diallo, I. Dia, Y. Ba. J. A. Ndione, A. A. Sall, A. Morse, A. Diop, M. Diallo M,
Statistical modeling of the abundance of vectors of West African Rift Valley fever in Barkédji,
Senegal. *PLoS One*. **12**, e114047 (2014).

431 21. D. Diallo, C. Talla, Y. Ba, I. Dia, A. A. Sall AA, M. Diall, Temporal distribution and spatial
432 pattern of abundance of the Rift Valley fever and West Nile fever vectors in Barkedji, Senegal. J
433 Vector Ecol. 2, 426-436 (2011).

434 22. M. J. Turell, C. A. Rossi, C. L. Bailey, Effect of extrinsic incubation temperature on the ability
435 of Aedes taeniorhynchus and Culex pipiens to transmit Rift Valley fever virus. *Am. J. Trop. Med.*436 *Hyg.* 134, 1211-1218 (1985).

437 23. J. F. Brubaker, M. J. Turell, Effect of environmental temperature on the susceptibility of Culex
438 pipiens (Diptera: Culicidae) to Rift Valley fever virus. *J. Med. Entomol.* 35, 918-921 (1998).

439 24. G. Lo Iacono, A. A. Cunningham, B. Bett, D. Grace, D. W. Redding, J. L. N. Wood,
440 Environmental limits of Rift Valley fever revealed using ecoepidemiological mechanistic models.
441 *Proc. Natl. Acad. Sci. U. S. A.* **115**, E7448-E7456 (2018).

442 25. R. Métras, M. Baguelin, W. J. Edmunds, P. N. Thompson, A. Kemp, D. U. Pfeiffer, L. M. 443 Collins, R. G. White RG. Transmission potential of Rift Valley fever virus over the course of the 444 2010 epidemic in South Africa. *Emerg. Infect. Dis.* **6**, 916-924 (2013).

445 26. H. J. Esser, R. Mögling, N. B. Cleton, H. van der Jeugd, H. Sprong, A. Stroo, M. P. G. 446 Koopmans, W. F. de Boer, C. B. E. M. Reusken, Risk factors associated with sustained 447 circulation of six zoonotic arboviruses: a systematic review for selection of surveillance sites in 448 non-endemic areas. *Parasit. Vectors.* **12**, 265 (2019).

449 27. C. M. Barker, T. Niu, W. K. Reisen, D. M. Hartley, Data-Driven Modeling to Assess 450 Receptivity for Rift Valley Fever Virus. *PLoS Negl. Trop. Dis.* **7**, e2515 (2013).

451 28. M. L. Danzetta, R. Bruno, F. Sauro, F. Savini, P. Calistri P, Rift Valley fever transmission 452 dynamics described by compartmental models. *Prev. Vet. Med.* **134**, 197–210 (2010).

453 29. T. Lernout, E. Cardinale, M. Jego, P. Desprès, L. Collet, B. Zumbo, E Tillard, S. Girard, L.
454 Filleul, Rift Valley Fever in Humans and Animals in Mayotte, an Endemic Situation? *PLoS ONE*.
455 8, e74192 (2013).

456 30. Cavalerie L, Charron MVP, Ezanno P, Dommergues L, Zumbo B, Cardinale E. A Stochastic 457 Model to Study Rift Valley Fever Persistence with Different Seasonal Patterns of Vector 458 Abundance: New Insights on the Endemicity in the Tropical Island of Mayotte. *PLoS One*. **10**, 459 e0130838 (2015).

31. Nicolas G, Chevalier V, Tantely LM, Fontenille D, Durand B. A Spatially Explicit
Metapopulation Model and Cattle Trade Analysis Suggests Key Determinants for the Recurrent
Circulation of Rift Valley Fever Virus in a Pilot Area of Madagascar Highlands. *PLoS Negl. Trop. Dis.* 8, e3346 (2014).

464 32. Gachohi JM, Njenga MK, Kitala P, Bett B. Modelling Vaccination Strategies against Rift Valley 465 Fever in Livestock in Kenya. *PLoS Negl. Trop. Dis.* **10** doi: 10.1371/journal.pntd.0005049 (2016).

33. T. Kimani, E. Schelling, B. Bett, M. Ngigi, T. Randolph, S. Fuhrimann, Public Health Benefits
from Livestock Rift Valley Fever Control: A Simulation of Two Epidemics in Kenya. *EcoHealth.* 13,
729–742 (2016).

34. World Health Organization, R&D Blueprint, Target Product Profiles for Rift Valley Fever Virus
Vaccines – version 3. Available at :

471 <u>https://www.who.int/docs/default-source/blue-print/call-for-comments/tpp-rift-valley-fever-</u>

472 <u>vaccines-draft3-0pc.pdf?sfvrsn=f2f3b314_2</u> Accessed 06 March 2020 (2019).

473 35. B. Dungu, B. A. Lubisi, T. Ikegami, Rift Valley fever vaccines : current and future needs. *Curr* 474 *Opin Virol.* **29**, 8-15 (2018).

47536. European Centre for Diseases Prevention and Control. Rift Valley fever outbreak in Mayotte,476France.RapidRiskassessment.Availableat477https://www.ecdc.europa.eu/sites/default/files/documents/RRA-Rift-Valley-fever-Mayotte-France-478March-2019.pdfAccessed 17 March 2020 (2019).

37. I. Scoones, K. Jones, G. Lo Iacono, D. W. Redding, A. Wilkinson, J. L. N. Wood, Integrative
modelling for One Health: pattern, process and participation. *Philos Trans R Soc Lond B Biol Sci.*372, 20160164 (2017).

38. B. H. Bird, D. A. Bawiec, T. G. Ksiazek, T. R. Shoemaker, S. T. Nichol, Highly sensitive and
broadly reactive quantitative reverse transcription-PCR assay for high-throughput detection of Rift
Valley fever virus. *J. Clin. Microbiol.* 45, 3506–3513 (2007).

39. H. Youssouf, M. Subiros, G. Dennetiere, L. Collet, L. Dommergues, A. Pauvert A, P.
Rabarison, C. Vauloup-Fellous, G. Le Godais, M. C. Jaffar-Bandjee, M. Jean, M. C. Paty, H.
Noel, S. Oliver, L. Filleul, C. Larsen, Rift Valley fever outbreak, Mayotte, France, 2018–2019. *Emerg. Infect. Dis.* Apr [08 March 2020] (2020).

489 40. J. Kortekaas, J. Kant, R. Vloet, C. Cêtre-Sossah, P. Marianneau, S. Lacote, A. C. Banyard, C.
490 Jeffries, M. Eiden, M. Groschup, S. Jäckel, E. Hevia, A. Brun, European ring trial to evaluate
491 ELISAs for the diagnosis of infection with Rift Valley fever virus. *J. Virol. Methods.* 1, 177-181
492 (2013).

493 41. Meteofrance. Donnees decadaires agrometeorologiques. Available at :

- 494 <u>https://donneespubliques.meteofrance.fr/?fond=produit&id_produit=113&id_rubrique=37</u>
- 495 Accessed on 23 September 2019 (2019).

496 43. Institut National de la statistique et des etudes economiques (Insee). Habitants a Mayotte.
497 Available at : <u>https://www.insee.fr/fr/statistiques/3286558#documentation</u>, Accessed on 05
498 October 2019 (2017).

499 43. A. Camacho, S. Funk, fitR: Tool box for fitting dynamic infectious disease models to time 500 series. R package version 0.1.

501 Ethics statement

502 The livestock data were collected under the under the Mayotte disease surveillance system 503 (Système d'Epidémiosurveillance Animale à Mayotte, SESAM) with the approval of the Direction 504 of Agriculture, Food and Forestry (DAAF) of Mayotte. For human data, according to French law, only "research involving a human being" (research defined by article L. 1121-1 and article R. 505 506 1121-1 of the Code de la santé publique) are compelled to receive the approval of ethics 507 committee. This study was based on anonymous data collected from health professionals for 508 public health purposes relating to the health surveillance mission entrusted to Santé publique 509 France by the French Law (article L. 1413-1 code de la santé publique). Therefore, the study did 510 not meet the criteria for gualifying a study "research involving a human being" and did not require 511 the approval of an ethics committee. Furthermore, as the data were anonymous, it did not require 512 an authorization of the French data protection authority (Commission Nationale informatique et 513 libertés).

514 Role of the funding sources

515 The funding sources have no role in study design; in the collection, analysis, and interpretation of

516 data; in the writing of the report; and in the decision to submit the paper for publication.

517 Declaration of interests

518 The authors declare no conflict of interest.

519 Funding sources

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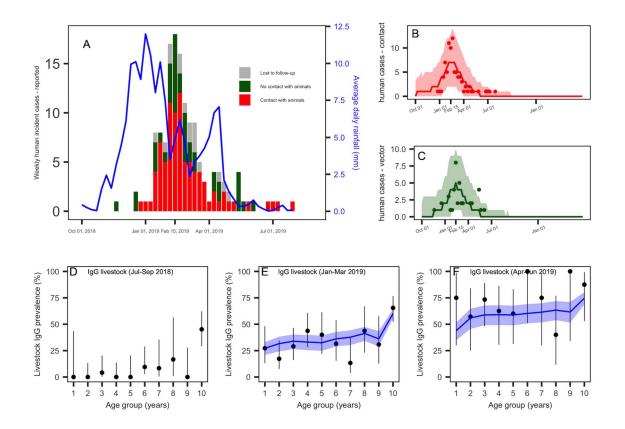


Figure 1AF. RVF epidemic data in humans and livestock, and model fit. (A) Weekly number 528 of reported human cases and average daily rainfall pattern (solid blue line). Human cases 529 reporting direct contact with animals or their products are presented in red (86 cases), those 530 531 reporting no prior contact with animals or their products are in green (41 cases), and lost to 532 follow-up are in grey (16 cases). (B) Predicted median (red solid line) and 95%Crl (red envelope) of the number of weekly reported human cases by direct contact, and weekly incident observed 533 cases by contact (red dots). (C) Predicted median (green solid line) and 95%Crl (green envelope) 534 535 of the number of weekly reported human cases by vector-mediated route, and weekly incident 536 observed cases with no prior contact with animals (green dots). (D) Quarterly age-stratified RVF 537 IgG seroprevalence in livestock for the trimesters July-September 2018 (N=173), (E) January-538 March 2019 (N=252), and (F) April-June 2019 (N=67). In (D), (E), (F), the black dots and vertical 539 black lines represent the observed age-stratified average IgG seroprevalence and their 95%CI. 540 The model predicted values are showed by the median (solid blue line) and 95%Crl (blue 541 envelopes).

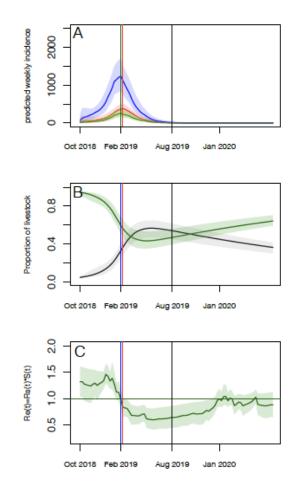
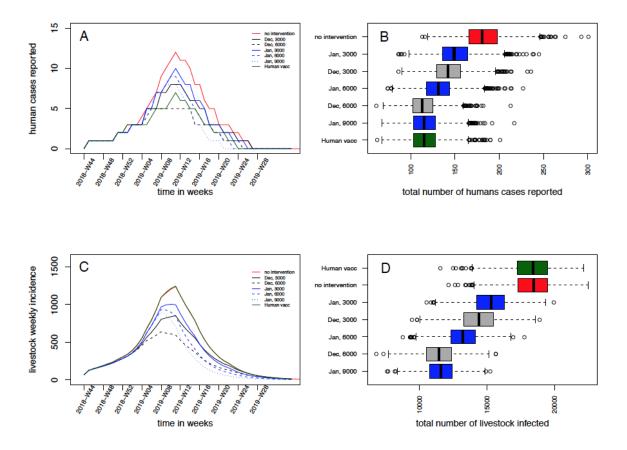


Figure 2AC. Model predictions over two rainy seasons (2018-2019 and 2019-2020): 542 epidemic curves, proportion of susceptible and immune livestock, and time-varying 543 544 effective reproduction ($R_{e}(t)$) number. (A) Predicted (reported and unreported) number of 545 infectious livestock (blue) and humans by direct contact (red), and vector-mediated route (green). 546 (B) Predicted median (solid lines) and 95.% Crl envelopes of the predicted proportion of Susceptible (green) and Immune (black) livestock over the course of the epidemic. (C) Values of 547 548 $R_{e}(t) = R_{s}(t) * S(t)$ over the course of the epidemic. In all panels, the vertical blue and red vertical 549 lines correspond to the predicted epidemic peaks in livestock and humans, respectively. The 550 vertical black line corresponds to the end of the fitting period (August 2019).



551 Figure 3AD. Figure 3AD. Impact of vaccination strategies on the epidemic size. (A) Median weekly number of predicted incident human cases, and corresponding (B) human epidemic size 552 (reported cases). (C) Median weekly number of predicted incident infected livestock, and 553 corresponding (D) total livestok epidemic size. In (A) and (C) the red solid line presents the 554 555 scenario with no intervention (Scenario 1); the black lines present vaccinations in December 2018 556 (black solid: 3,000 doses, dashed black: 6,000 doses) (Scenarios 2-3); the blue lines present the 557 vaccinations in January 2019 (blue solid: 3,000 doses, dashed blue: 6,000 doses; dotted blue: 558 9,000 doses) (Scenarios 4-6); the red line represent the human vaccination only (Scenario 7).

Table 1. Predicted epidemic sizes (total and reported cases) and post-epidemic prevalence in 560 med Reimansiandi livestock, for the different scenarios, the distortion were after the stock of the different scenarios, the distort of the different scenarios, the different scenario

	Epidemic size in a made available under a do bit no 4.6 international license rost-epidemic prevalence						
Scenarios	Livestock total	Humans total	Humans	Reported	Reported	Livestock	Humans
			reported	contact	vector		
1. No	18,461	9,566	181	111	70	53.92	3.73
intervention	(14,926-21,153)	(7,793-11,772)	(138-233)	(79-149)	(44-102)	(44.76-61.29)	(3.03-4.56)
Livestock vac	cination						
2. Dec 3,000	14,415	7,465	142	87	54	42.40	2.90
	(11,154-17,237)	(5,980-9,381)	(106-186)	(61-119)	(34-80)	(34.21-50.33)	(2.33-3.65)
3. Dec 6,000	11,447	5,936	113	69	43	34.17	2.31
	(8,863-14,046)	(4,683-7,674)	(83-151)	(46-97)	(26-67)	(27.66-41.15)	(1.82-2.99)
4. Jan, 3000	15,311	7,867	149	91	58	44.72	3.06
	(12,237-17,985)	(6,302-9,956)	(112-197)	(63-126)	(35-84)	(36.85-51.72)	(2.45-3.88)
5. Jan, 6000	13,229	6,872	131	80	51	38.87	2.68
	(10,545-15,693)	(5,470-8,752)	(96-175)	(55-112)	(32-77)	(32.21-45.44)	(2.13-3.41)
6. Jan, 9000	11,573	6,014	115	90	44	34.27	2.34
	(9,296-13,784)	(4,665-7,992)	(83-158)	(47-101)	(27-68)	(28.52-39.86)	(1.82-3.12)
Human vaccir	nation	•					
7. Dec, 50 %	18,436	6,032	115	70	44	53.79	2.35
	(14,987-21,099)	(4,708-7,942)	(82-157)	(46-102)	(26-67)	(45.05-61.20)	(1.83-3.10)