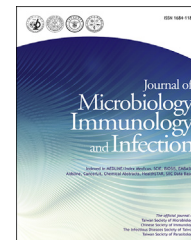


Available online at www.sciencedirect.com

ScienceDirect

journal homepage: www.e-jmii.com

Correspondence

Duration of serum neutralizing antibodies for SARS-CoV-2: Lessons from SARS-CoV infection

Dear editor,

The 2019 novel coronavirus (2019-nCoV, now designated as severe acute respiratory syndrome coronavirus 2, SARS-CoV-2) outbreak has posed a severe threat to public health. The epidemiological and clinical features of the novel CoV infection have been elucidated by some studies. As the number of recovered patients with coronavirus disease 2019 (COVID-19) continues to be increasing, a critical issue has aroused wide concern: will patients recovering from COVID-19 be reinfected? Answering the question is of great help for the follow-up of convalescent patients and the control of epidemic.

The illustrated pathogenicity of SARS-CoV-2 seems to be similar to SARS-CoV in some ways. Besides, a recent study showed that neutralizing antibody from a convalescent SARS patient could block the SARS-CoV-2 from entering into target cells *in vitro*,¹ which implies the potential cross-protective epitope between the two viruses. Thus, the potential immune protection against reinfection with SARS-CoV-2 may share some common features with convalescent SARS-CoV.

Protection specific Abs, including immunoglobulin G (IgG) Abs and neutralizing Abs (NAbs), are produced by B cells after infection with the virus, which can block the virus from entering the host cells and defense against viral reinfection. A cohort study of convalescent SARS-CoV patients (56 cases, from Beijing hospital of Armed Forces Police, China) revealed that the specific IgG Abs and NAbs were highly correlated,² peaking at month 4 after the onset of disease and decreasing gradually thereafter (Fig. 1). Since titers decreased markedly after month 16, IgG Abs and NAbs remained detectable in all patients throughout 2 years follow-up, except for at the last visit (at month 24) in which 11.8% samples turned to negative. Then Cao et al. extended the follow-up time to 3 years, IgG Abs and NAbs were detectable in 74.2% and 83.9%, respectively, at month 36.³ Another study (176 cases, Shanxi 7 designated SARS

hospitals, China) showed a similar result, up to 11.8% serum samples were IgG Abs positive at day 7 after the onset of symptom of SARS-CoV.⁴ The positive rate increased gradually, reached 100% at day 90, and remained unchanged until day 200. About 93.9% and 89.6% of these patients, respectively, were detectable after 1 and 2 years. Notably, 3 years later, 50% of the convalescent patients still had SARS-CoV specific IgG Abs. The longest follow-up study (23 cases, Beijing, China) showed that only 8.7% patients of recovered patients sustained low levels of specific IgG Ab to SARS-CoV.⁵ These finds suggested that the immune responses of specific Abs were maintained in more than 90% of recovered SARS-CoV patients for 2 years.

Since the SARS-CoV epidemic 17 years ago, specific IgG Abs to SARS-CoV may have been waned in most surviving patients, which could not be sufficient to protect against challenge with SARS-CoV-2 infection. However, previous experimental SARS-CoV vaccines and neutralizing antibody could be a novel preventive and therapeutic option for COVID-19. Besides, not all (only 11.8% at day 7 and reached

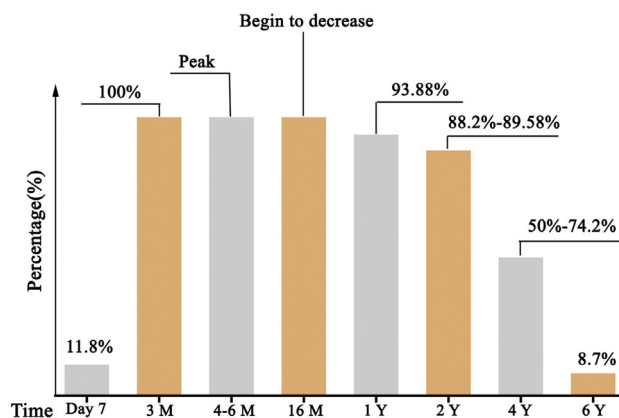


Fig. 1. The percentage of patients who expressed specific IgG Abs/NAbs against SARS-CoV in recovered patients²⁻⁴.

<https://doi.org/10.1016/j.jmii.2020.03.015>

1684-1182/Copyright © 2020, Taiwan Society of Microbiology. Published by Elsevier Taiwan LLC. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

100% at day 90) patients acquire specific SARS-CoV Abs in the early period after recovery, which highlights the importance of the detection of Abs titers for convalescent COVID-19 patients. Otherwise, these patients with low titers of Abs may not efficient for the clearance of SARS-CoV-2. These experience from SARS-CoV are expected to have some implications for the treatment, management and surveillance of SARS-CoV-2 patients.

Funding

The research was supported by National Natural Science Foundation of China (No. 81873451).

Acknowledgement

The correspondence author, doctor You, participate in the fight against the SARS-CoV-2 on the frontline. The authors gratefully acknowledge thousands of unsung heroes in the fight against the epidemic of COVID-19.

References

1. Markus H, Hannah KW, Nadine K, Marcel M, Christian D, Stefan P, et al. The novel coronavirus 2019 (2019-nCoV) uses the SARS-coronavirus receptor ACE2 and the cellular protease TMPRSS2 for entry into target cells. *bioRxiv* 2020. <https://doi.org/10.1101/2020.01.31.929042>. published online Jan 31.
2. Liu W, Fontanet A, Zhang PH, Zhan L, Xin ZT, Baril L, et al. Two-Year prospective study of the humoral immune response of patients with severe acute respiratory syndrome. *J Infect Dis* 2006; **193**:792–5.
3. Cao WC, Liu W, Zhang PH, Zhang F, Richardus JH. Disappearance of antibodies to SARS-associated coronavirus after recovery. *N Engl J Med* 2007; **357**:1162–3.
4. Wu LP, Wang NC, Chang YH, Tian XY, Na DY, Zhang LY, et al. Duration of antibody responses after severe acute respiratory syndrome. *Emerg Infect Dis* 2007; **13**(10):1562–4.
5. Tang F, Quan Y, Xin ZT, Wrammert J, Ma MJ, Lv H, et al. Lack of peripheral memory B cell responses in recovered patients with severe acute respiratory syndrome: a six-year follow-up study. *J Immunol* 2011; **186**:7264–8.

Qingqing Lin¹

Department of Hematology, The First Affiliated Hospital,
College of Medicine, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China
Institute of Hematology, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China

Li Zhu¹

Department of Hematology, The First Affiliated Hospital,
College of Medicine, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China
Malignant Lymphoma Diagnosis and Therapy Center, The
First Affiliated Hospital, College of Medicine, Zhejiang
University, Hangzhou, 310003, Zhejiang, PR China
Institute of Hematology, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China

Zuowei Ni¹

Department of Hematology, The First Affiliated Hospital,
College of Medicine, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China
The First Affiliated Hospital, College of Medicine, Zhejiang
University, Hangzhou, 310003, Zhejiang, PR China

Haitao Meng**

Department of Hematology, The First Affiliated Hospital,
College of Medicine, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China
Malignant Lymphoma Diagnosis and Therapy Center, The
First Affiliated Hospital, College of Medicine, Zhejiang
University, Hangzhou, 310003, Zhejiang, PR China
Institute of Hematology, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China

Liangshun You*

Department of Hematology, The First Affiliated Hospital,
College of Medicine, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China
Malignant Lymphoma Diagnosis and Therapy Center, The
First Affiliated Hospital, College of Medicine, Zhejiang
University, Hangzhou, 310003, Zhejiang, PR China
Institute of Hematology, Zhejiang University, Hangzhou,
310003, Zhejiang, PR China

**Corresponding author. Department of Hematology, The
First Affiliated Hospital, College of Medicine, Zhejiang
University, Hangzhou 310003, Zhejiang, PR China.

*Corresponding author. Department of Hematology, The
First Affiliated Hospital, College of Medicine, Zhejiang
University, 79# Qingchun Road, Hangzhou, 310003, PR
China.

E-mail address: youliangshun@zju.edu.cn (L. You)

12 March 2020

Available online ■ ■ ■

¹ Co-first author.